

# Raman spectroscopy: Variants and potentials

Hiro-o Hamaguchi

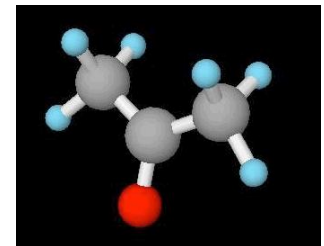
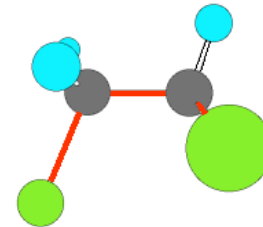
National Chiao Tung University, Taiwan  
The University of Tokyo, Japan  
Waseda University, Japan

# Raman spectroscopy: Variants and potentials

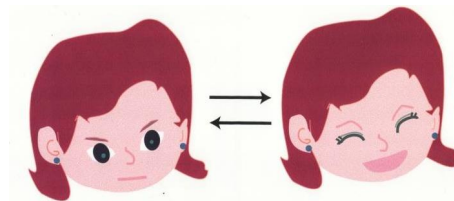
- Raman brothers,  
Raman, CARS/SRS, and SERS/TERS  
What do they look at?  
Hamaguchi's score sheet for Raman brothers



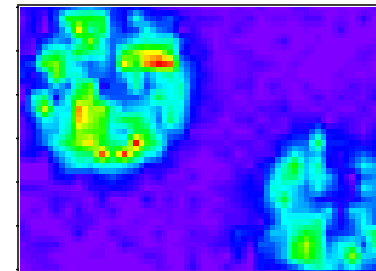
- Raman spectroscopy in Japan  
Discovery of rotational isomerism  
Normal coordinate analysis



- Time-resolved Raman spectroscopy  
Dynamic polarization model of  
chemical reactions

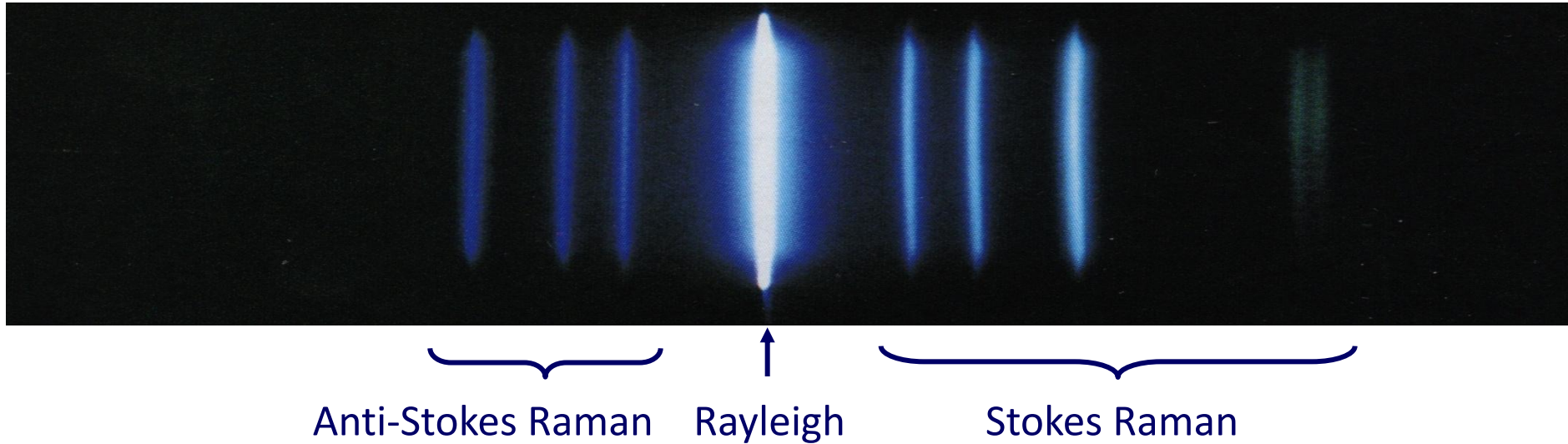


- Raman microspectroscopy of living cells



- Tailor-made Raman spectroscopy

# Raman Spectroscopy



Raman scattering :

Inelastic light scattering discovered by C. V. Raman in 1928

Raman spectroscopy:

Spectroscopy utilizing Raman scattering



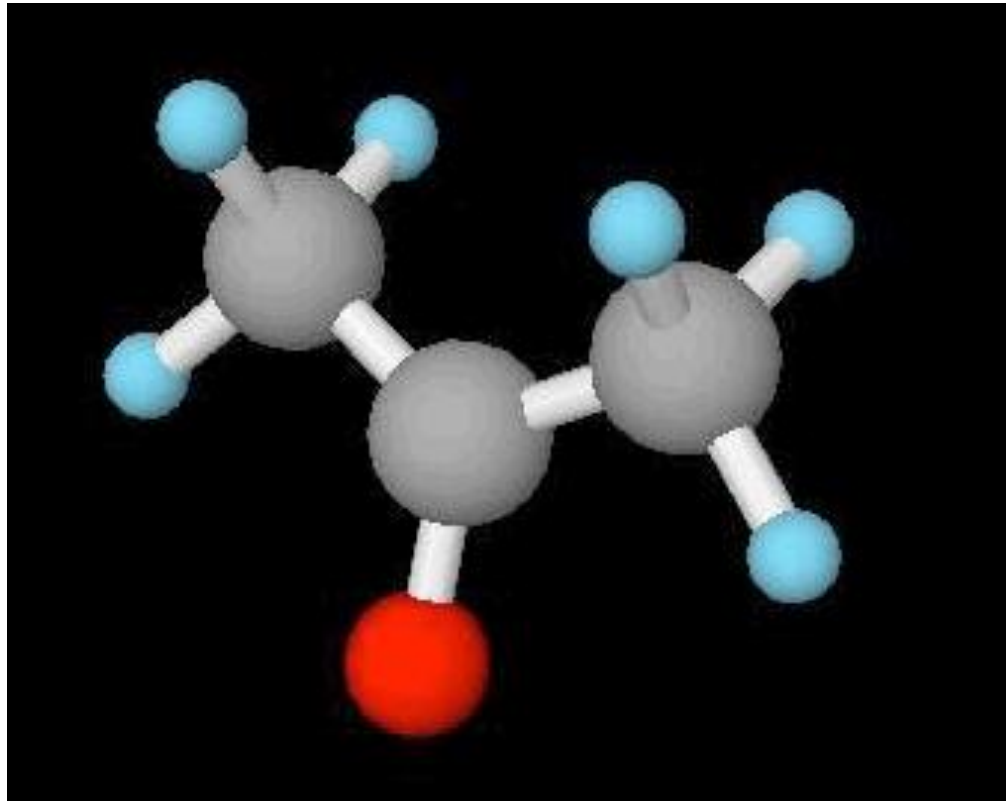
C. V. Raman (1888-1970)

# Raman Brothers: What do they look at?

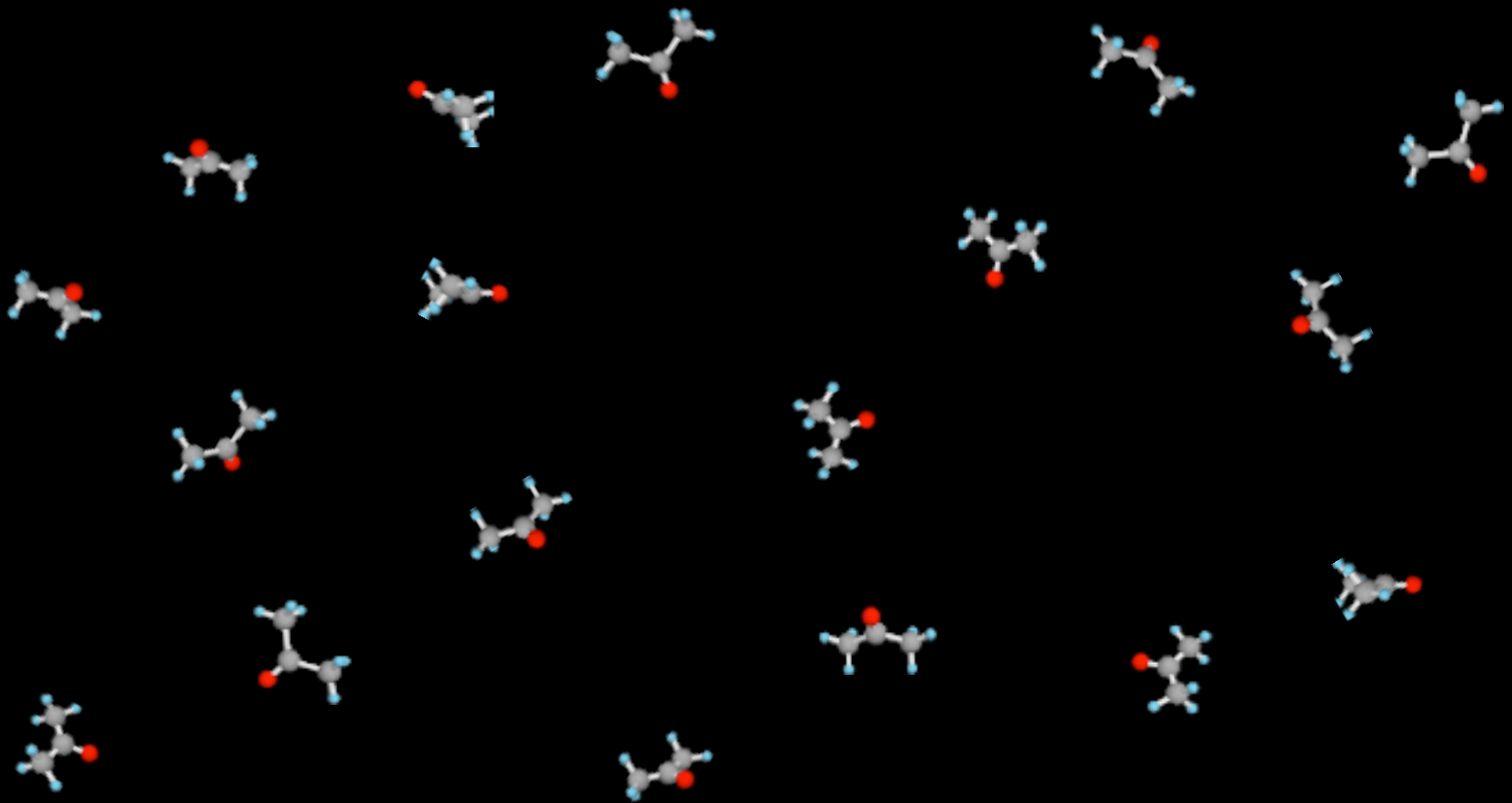


# Normal Mode of Vibration:

The Totally Symmetric C-C Stretch Mode of an Acetone Molecule

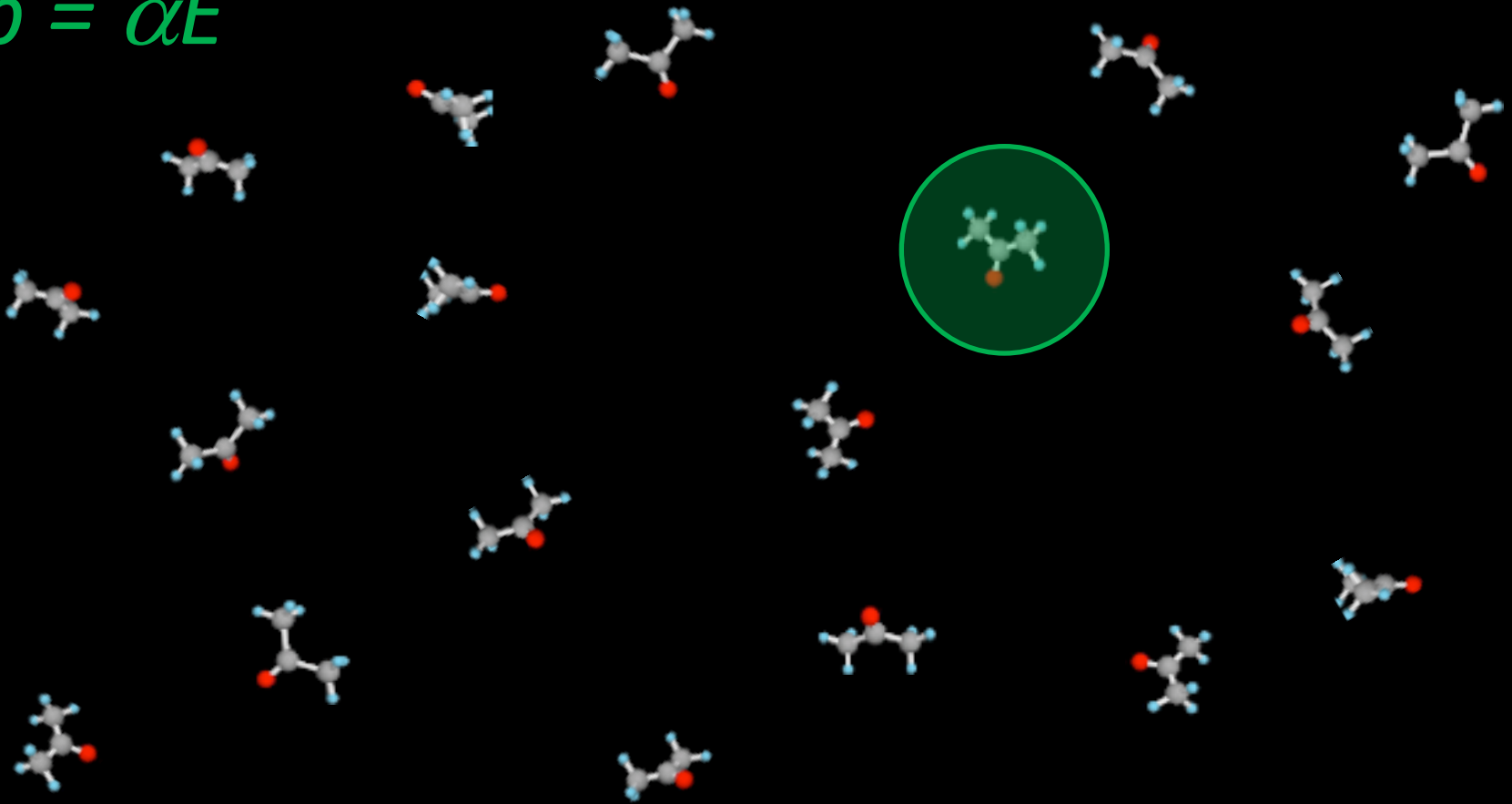


# Vibrational Motions of Ensembles of Acetone Molecules

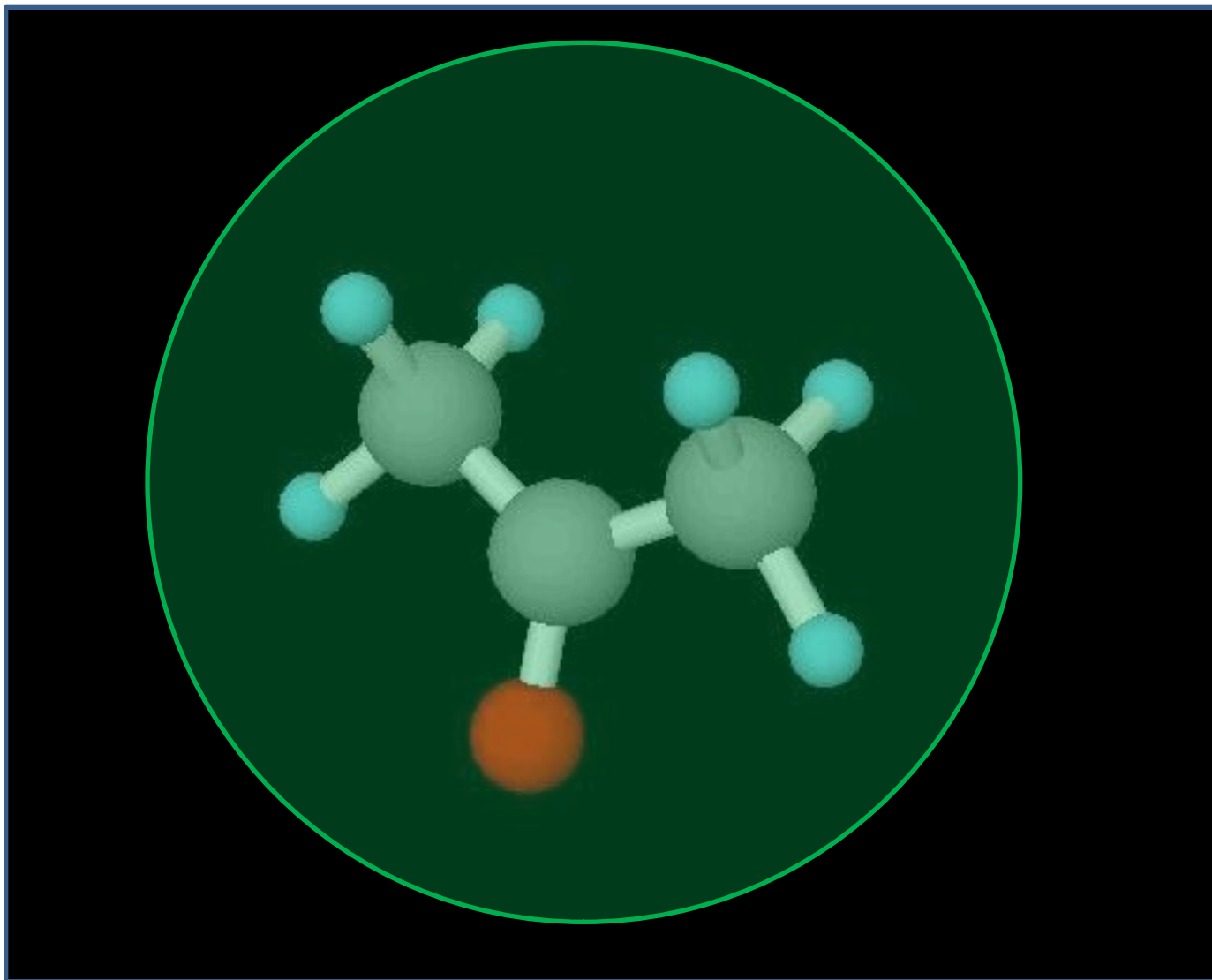


# What Does Raman look at?

$$p = \alpha E$$



# Raman Looks at a Normal Mode of Vibration.

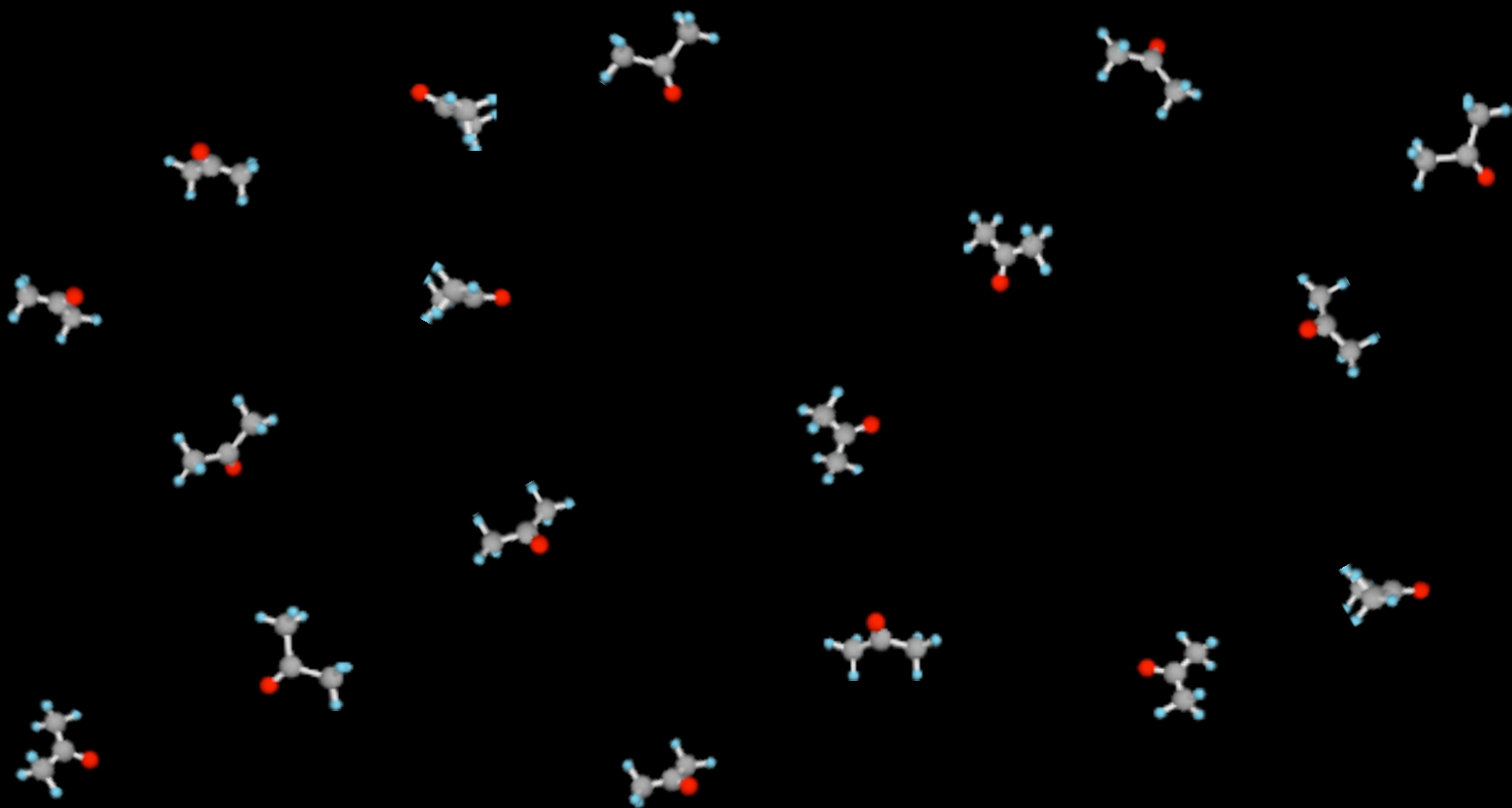




# Raman Gives a Molecular Fingerprint.

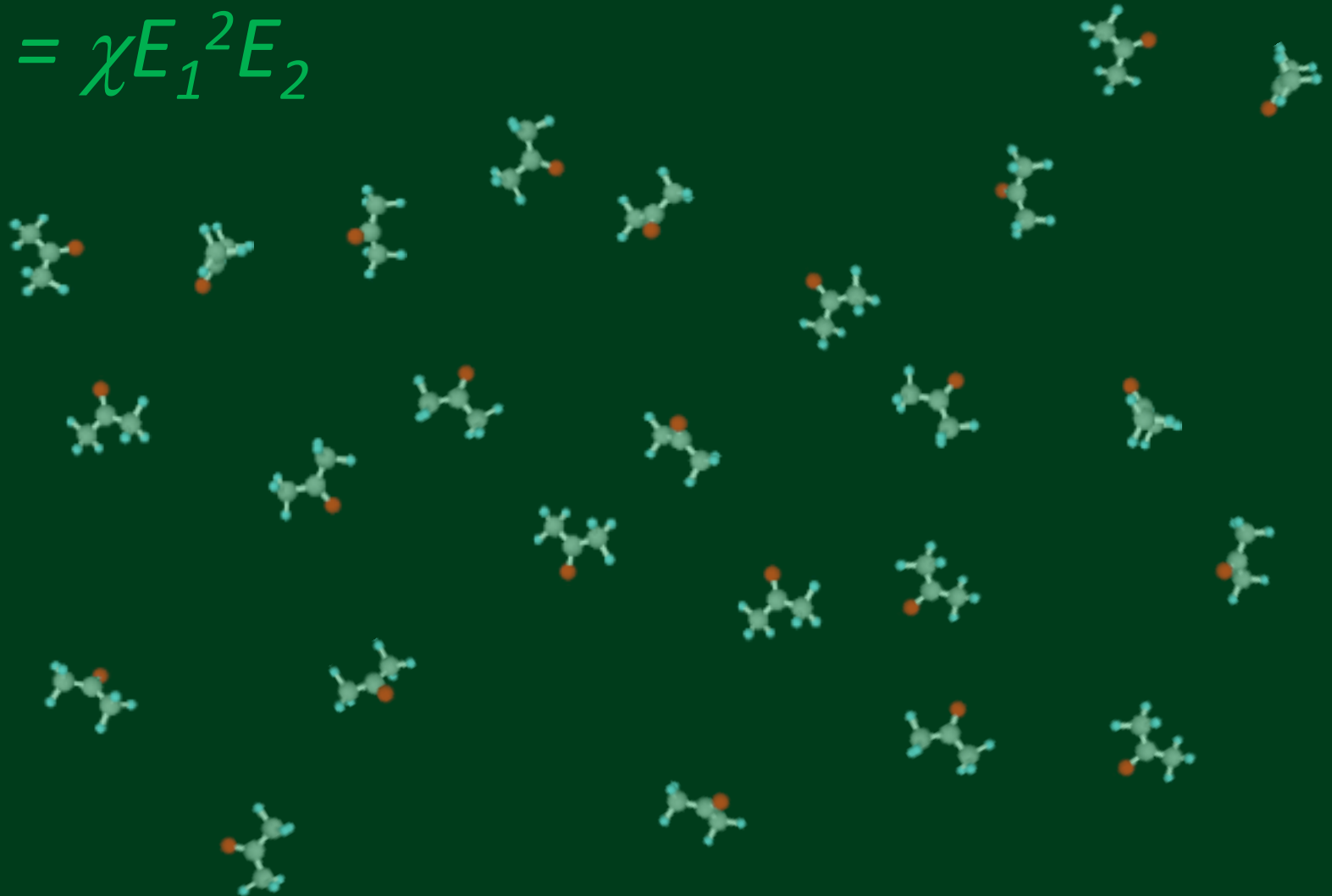


# What Does *CARS/SRS* Look at?



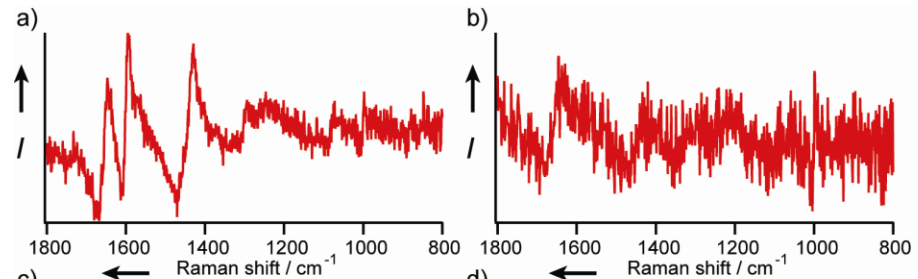
# CARS/SRS Looks at a Vibrational Coherence

$$p = \chi E_1^2 E_2$$

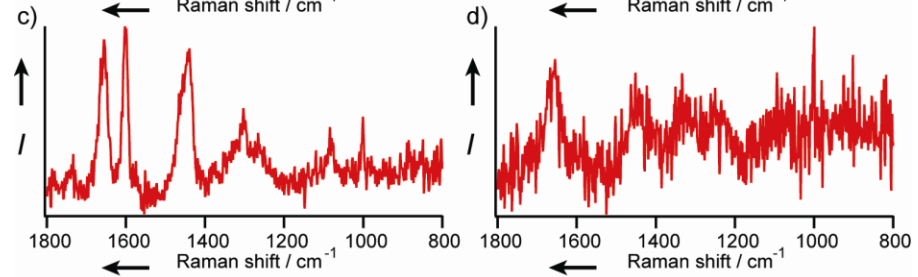


# CARS Can Give a Molecular Fingerprint After Some Mathematical Treatments.

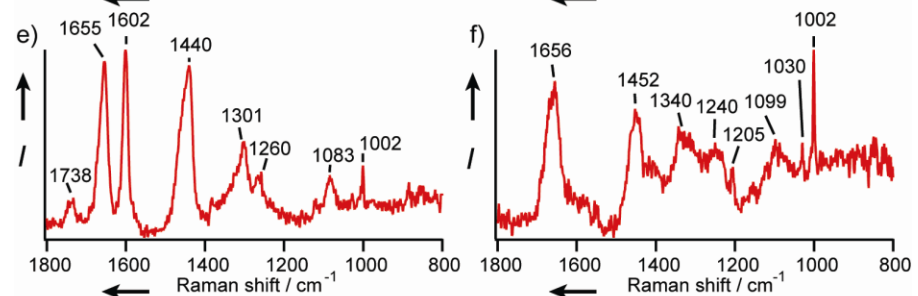
CARS spectra



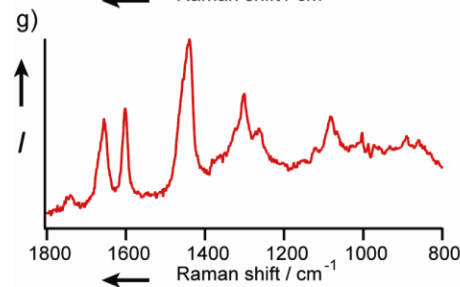
$\text{Im}\chi_3$  spectra by MEM



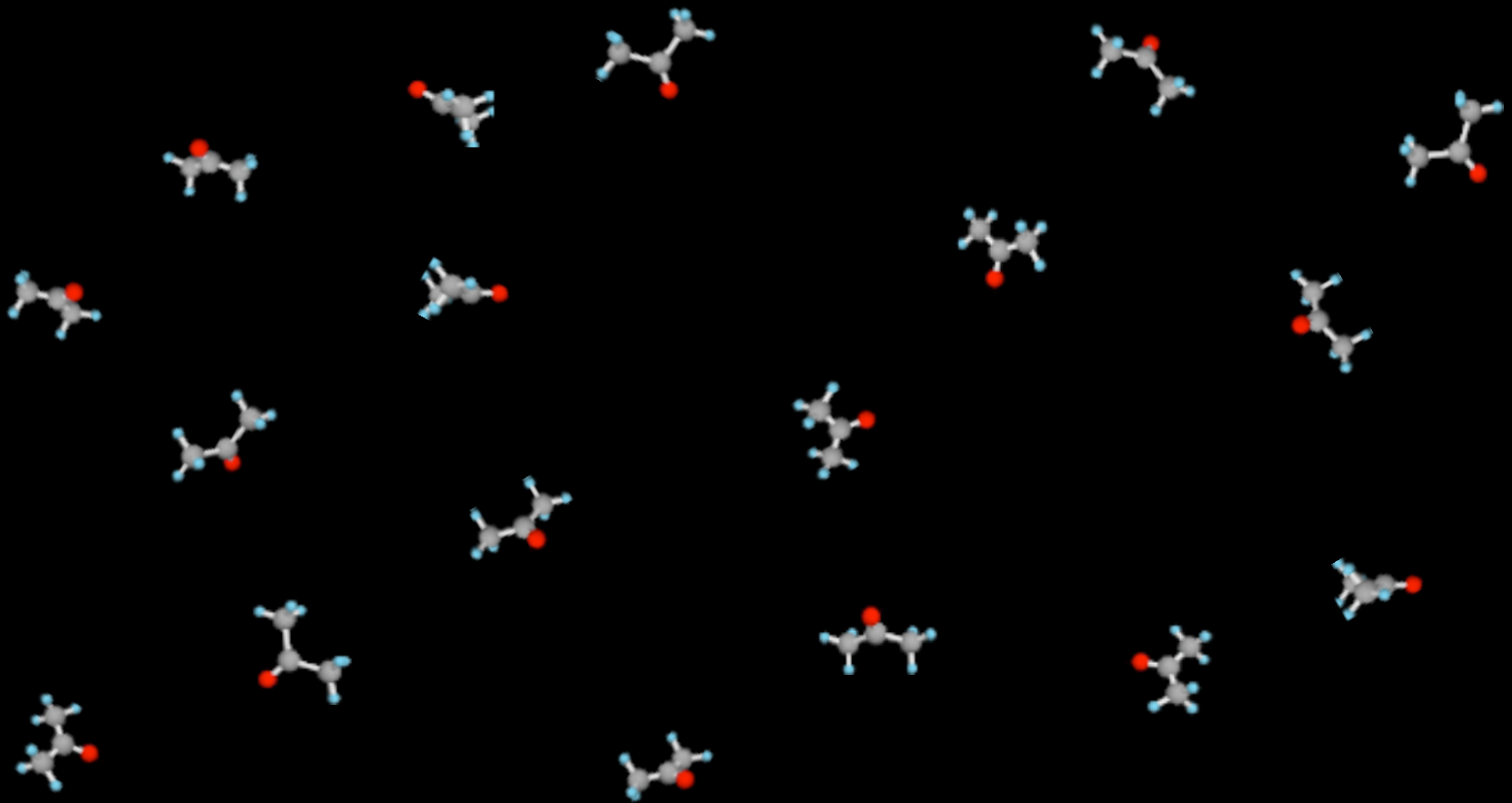
$\text{Im}\chi_3$  spectra after SVD



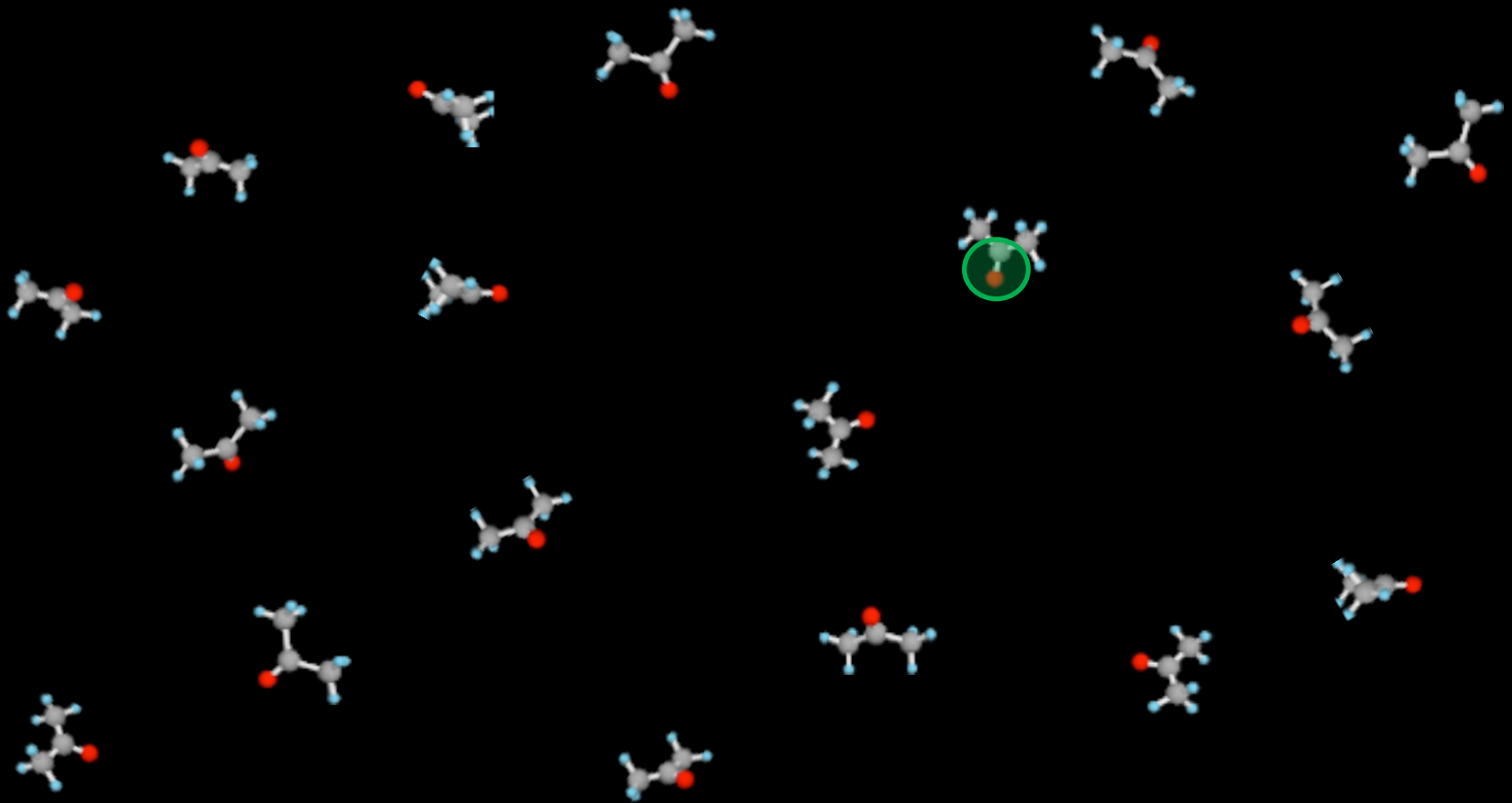
Raman spectra



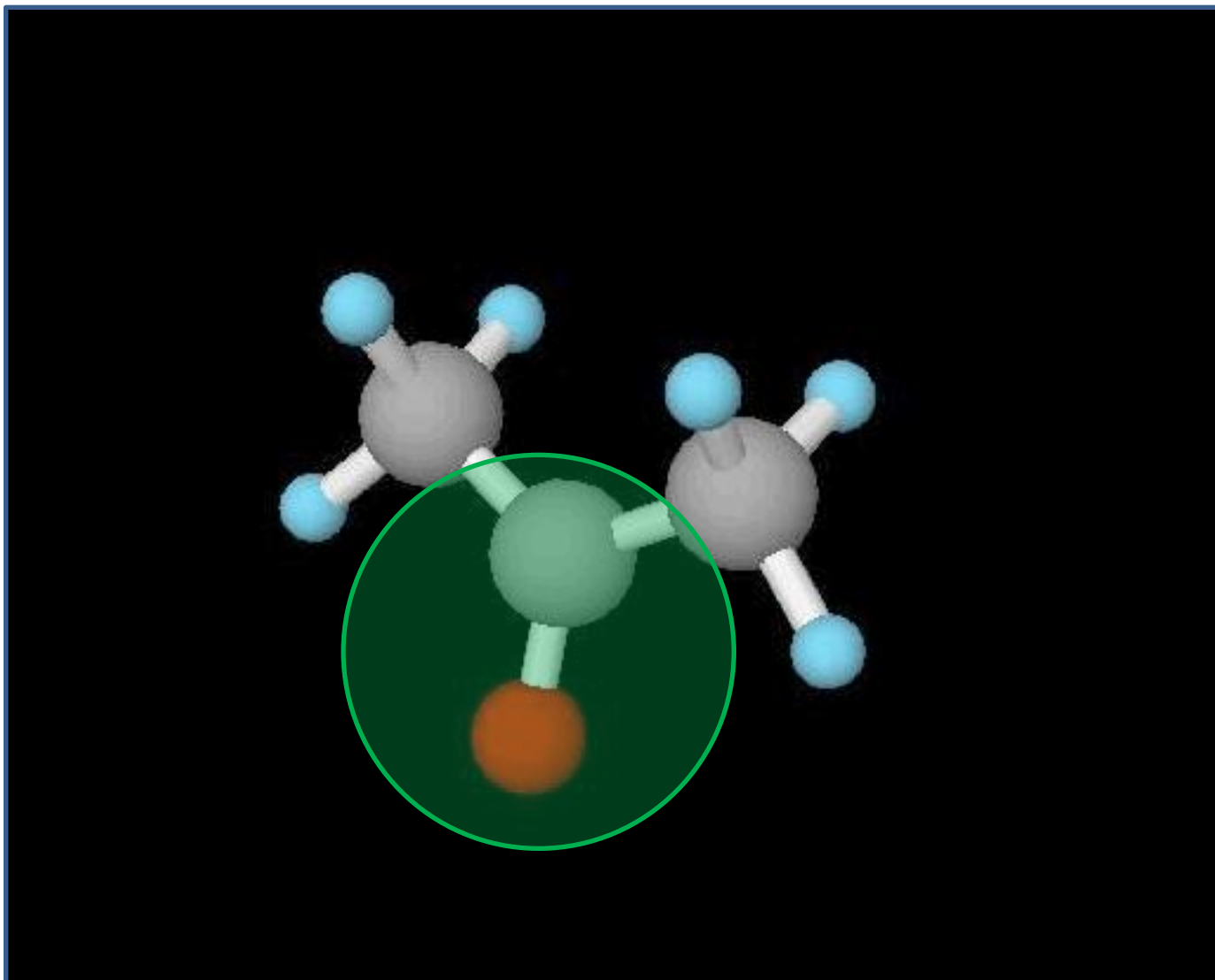
# Vibrational Motions of Ensembles of Acetone Molecules



# What Does SERS/TERS Look at?



SERS/TERS is Likely to Look at a Local Molecular Motion  
(May not be a Normal Mode)



# SERS/TERS May not Give a Full Molecular Fingerprint.



<http://dailynewsagency.com>



# Hamaguchi's Score Sheet for Ramans

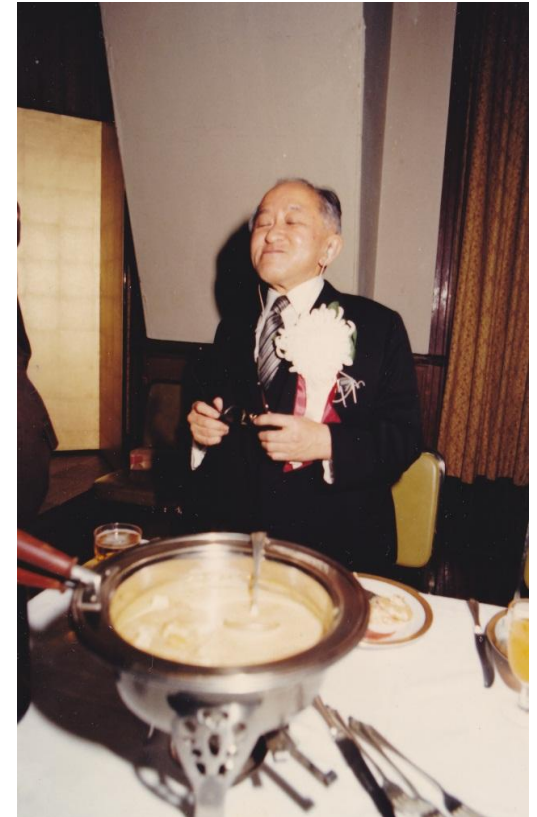
	Raman	CARS/SRS	SERS/TERS
Selection rule	Established	Established for homogeneous systems	Dependence on individual molecular orientation?
Polarization rule	Established	Established for homogeneous systems High polarization capabilities	Dependence on individual molecular orientation?
Information content	Molecular Fingerprint	Molecular fingerprint after mathematics	Molecular Fingerprint?
Sensitivity	Ensemble	Ensemble	Single molecule
Spatial resolution	$0.61\lambda/NA$ (500 nm)	$0.61\lambda/NA\sqrt{3}$ (300 nm)	Tip size (10 nm)
Time resolution	ps	ps	ps
Experimental difficulty	Fluorescence interference	Phase match/mismatch	Tip/substrate dependence

○ Raman is HH's first choice for advanced applications.

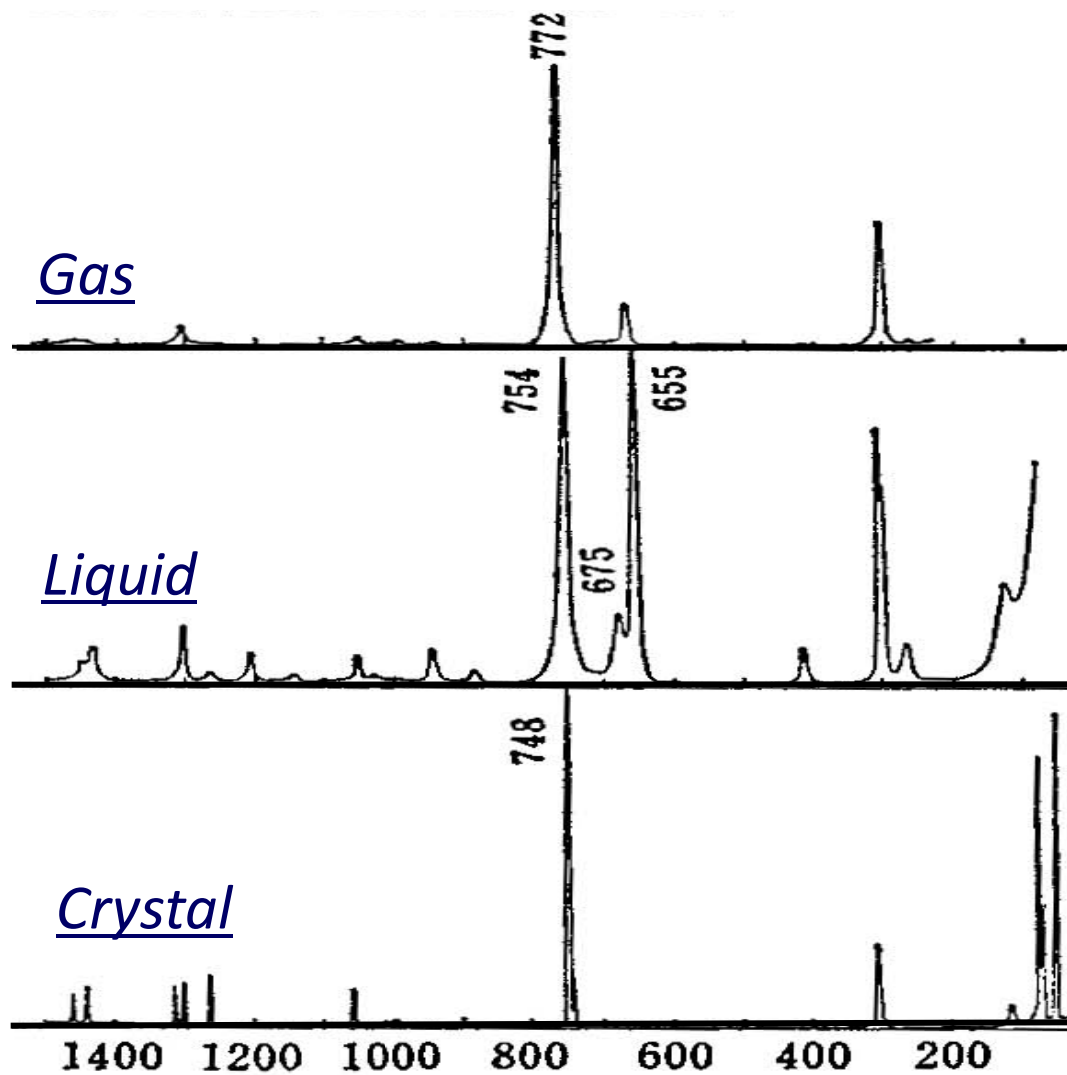
○ CARS/SRS surpasses Raman for its polarization capabilities.

○ SERS/TERS excels in sensitivity and spatial resolution. Its information content yet to be clarified.

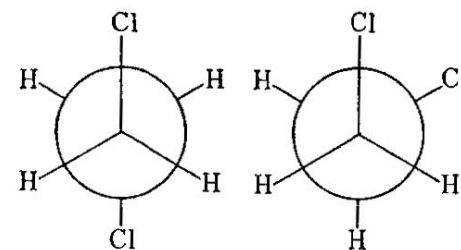
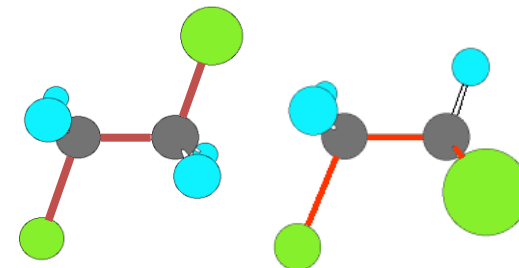
# Raman Spectroscopy at Tokyo



# Discovery of the Rotational Isomerism



Trans   Gauche



S. Mizushima  
(1899-1983)



*Raman Spectra are  
letters from the  
molecule*

*Vibrational Raman  
spectra are  
molecular  
fingerprints*

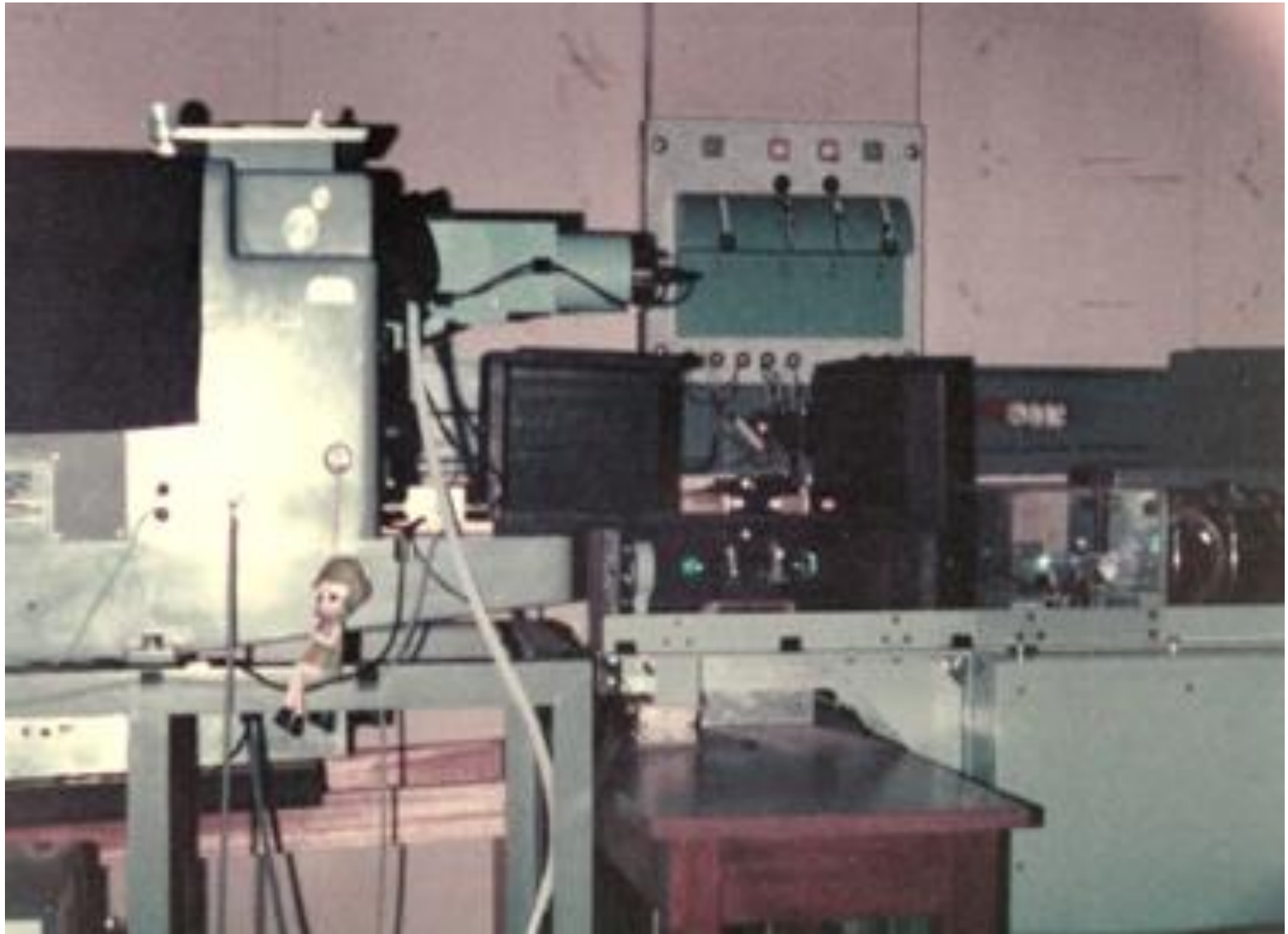
# T. Shimanouchi, Tables of Molecular Vibrational Frequencies, NSRDS-NBS 39.

Molecule: **1,2-Dichloroethane**  $\text{CH}_2\text{ClCH}_2\text{Cl}$  (trans form)  
Symmetry  $\text{C}_{2h}$  Symmetry number  $\sigma = 2$

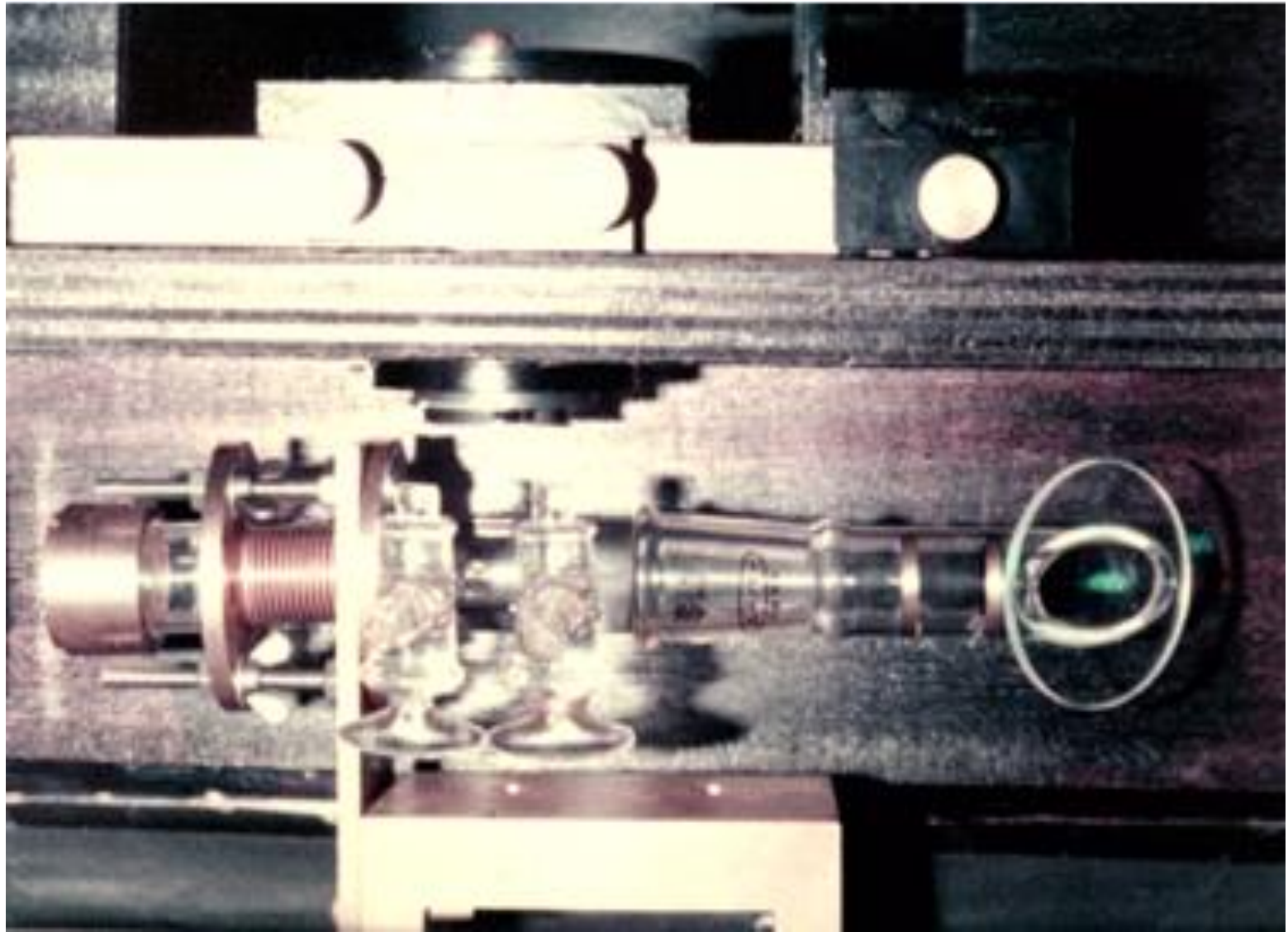
No. 160

Sym. species	No.	Approximate type of mode	Selected value of frequency	Infrared	Raman	Comments
				$\text{cm}^{-1}$ (Gas)	$\text{cm}^{-1}$ (Liquid)	
$a_g$	$\nu_1$	$\text{CH}_2$ s-stretch . . . . .	2957 D	ia	2957 (10) p	
	$\nu_2$	$\text{CH}_2$ scis . . . . .	1445 C	ia	1445 (4b) dp	
	$\nu_3$	$\text{CH}_2$ wag . . . . .	1304 C	ia	1304 (6) p	
	$\nu_4$	CC stretch . . . . .	1052 C	ia	1052 (4) p	
	$\nu_5$	CCl stretch . . . . .	754 C	ia	754 (10b) p	
	$\nu_6$	CCCl deform . . . . .	300 C	ia	300 (8) p	
$a_u$	$\nu_7$	$\text{CH}_2$ a-stretch . . . . .	3005 D	3005 W (liquid)	ia	SF (gauche $\nu_1$ , gauche $\nu_{11}$ ).
	$\nu_8$	$\text{CH}_2$ twist . . . . .	1123 B	1122.5 W	ia	
	$\nu_9$	$\text{CH}_2$ rock . . . . .	773 B	772.5 M	ia	
	$\nu_{10}$	Torsion . . . . .	123 C	123 M	ia	
$b_g$	$\nu_{11}$	$\text{CH}_2$ a-stretch . . . . .	3005 D	ia	3005 (8b) dp	
	$\nu_{12}$	$\text{CH}_2$ twist . . . . .	1264 C	ia	1264 (3) dp	
	$\nu_{13}$	$\text{CH}_2$ rock . . . . .	989 C	ia	989 (2) p	
$b_u$	$\nu_{14}$	$\text{CH}_2$ s-stretch . . . . .	2983 C	2983.3 M	ia	
	$\nu_{15}$	$\text{CH}_2$ scis . . . . .	1461 A	1460.6 S	ia	
	$\nu_{16}$	$\text{CH}_2$ wag . . . . .	1232 B	1232.3 S	ia	
	$\nu_{17}$	CCl stretch . . . . .	728 C	728.3 VS	ia	
	$\nu_{18}$	CCCl deform . . . . .	222 C	222.3 W	ia	

# High Resolution Raman Spectrometer (1972)

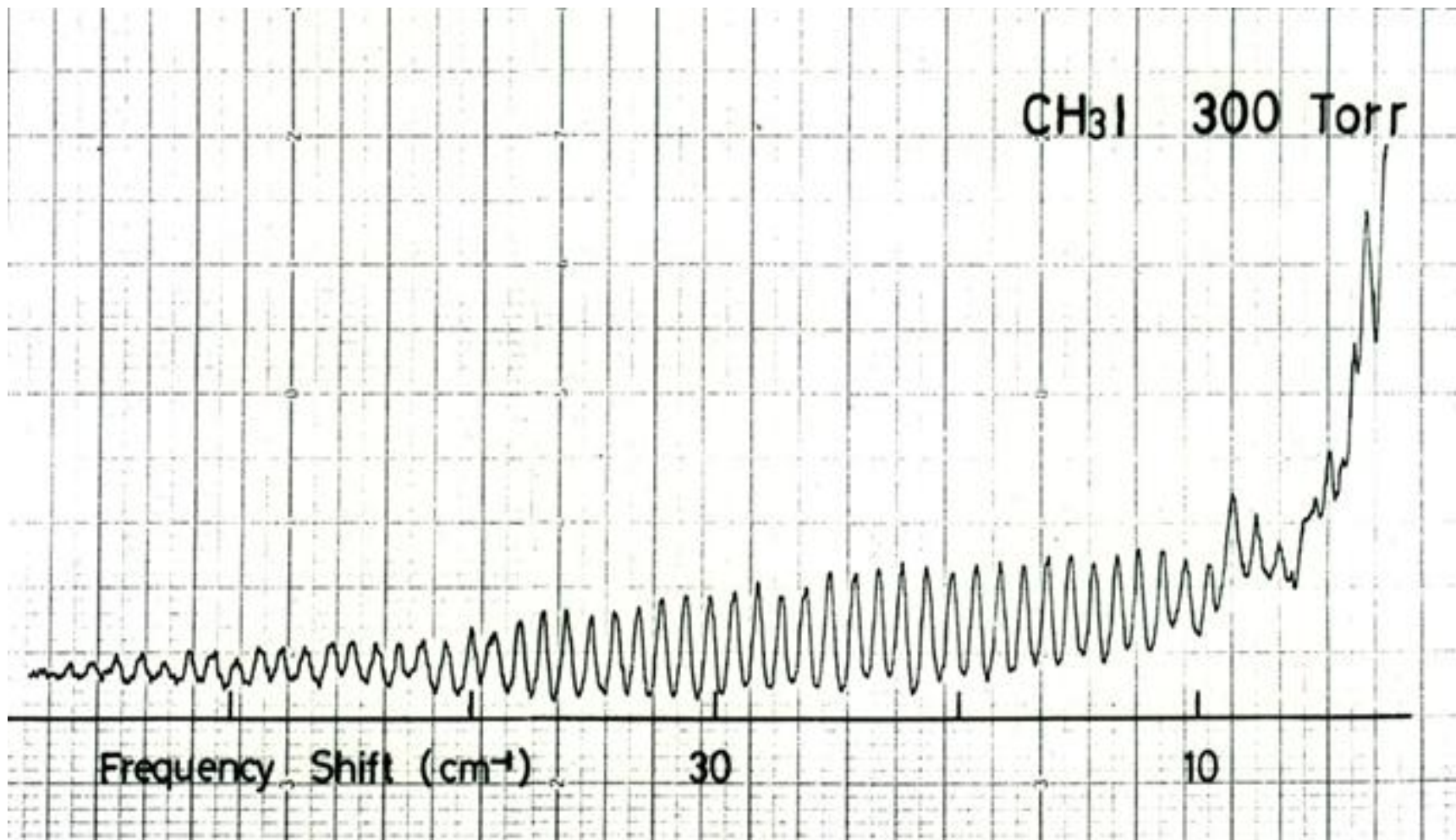


# Raman Gas Cell in Laser Cavity (1972)





# Raman Spectrum of CH<sub>3</sub>I (1972)



# Resonance Raman scattering

## RESONANCE RAMAN SPECTROSCOPIC STUDY ON IODINE IN VARIOUS ORGANIC SOLVENTS: SPECTROSCOPIC CONSTANTS AND HALFBAND WIDTHS OF THE I<sub>2</sub> VIBRATION\* \*\*

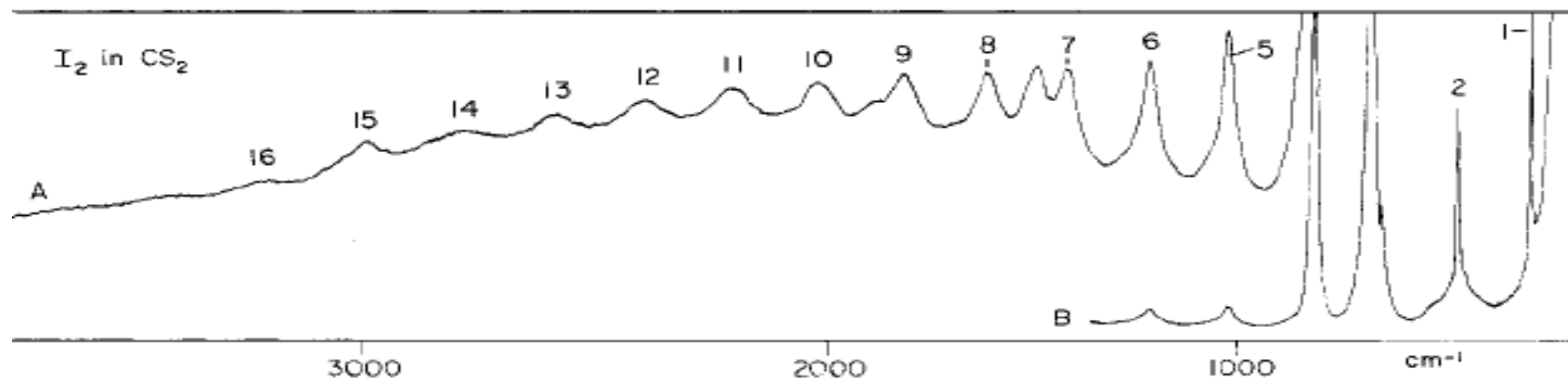
W. KIEFER†

*Sektion Physik der Universität München, Lehrstuhl J. Brandmüller,  
D-8 München 40, Germany*

and

H. J. BERNSTEIN

*Division of Chemistry, National Research Council of Canada, Ottawa, Ontario, Canada*



# Theory of Resonance Raman Scattering

## Albrecht's vibronic theory of resonance Raman Scattering

A. C. Albrecht, J. Chem. Phys. 34, 1476 (1961).

$$a_{\rho\sigma} \sim A + B$$

A. C. Albrecht  
(1927-2002)



A term: Franck-Condon term

Totally symmetric modes

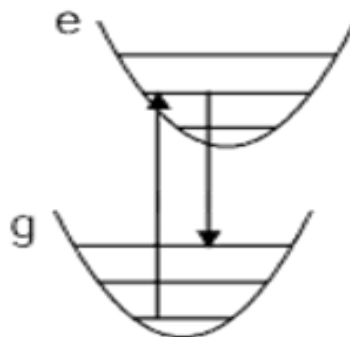
High overtones

B term: Vibronic coupling

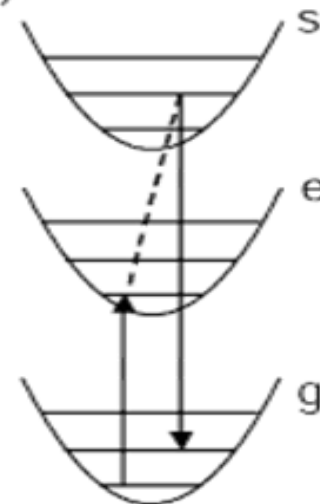
Non-totally symmetric modes

No high overtones

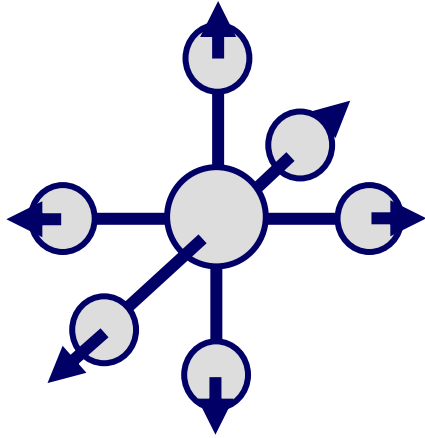
(a)



(b)



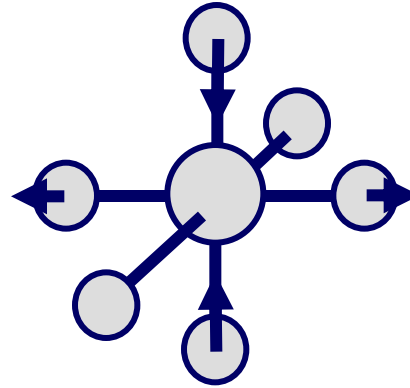
# Raman Active Vibrations of $MX_6$ Octahedral Complexes



$\nu_1$   
( $a_{1g}$ )

$$a_{1g} \sim \begin{pmatrix} 1 & 0 & 0 \\ 0 & 1 & 0 \\ 0 & 0 & 1 \end{pmatrix}$$

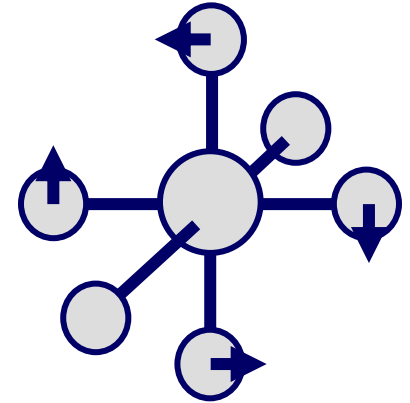
$\rho=0$



$\nu_2$   
( $e_g$ )

$$e_g \sim \begin{pmatrix} 1 & 0 & 0 \\ 0 & -1 & 0 \\ 0 & 0 & 0 \end{pmatrix}$$

$\rho=0.75$



$\nu_5$   
( $t_{2g}$ )

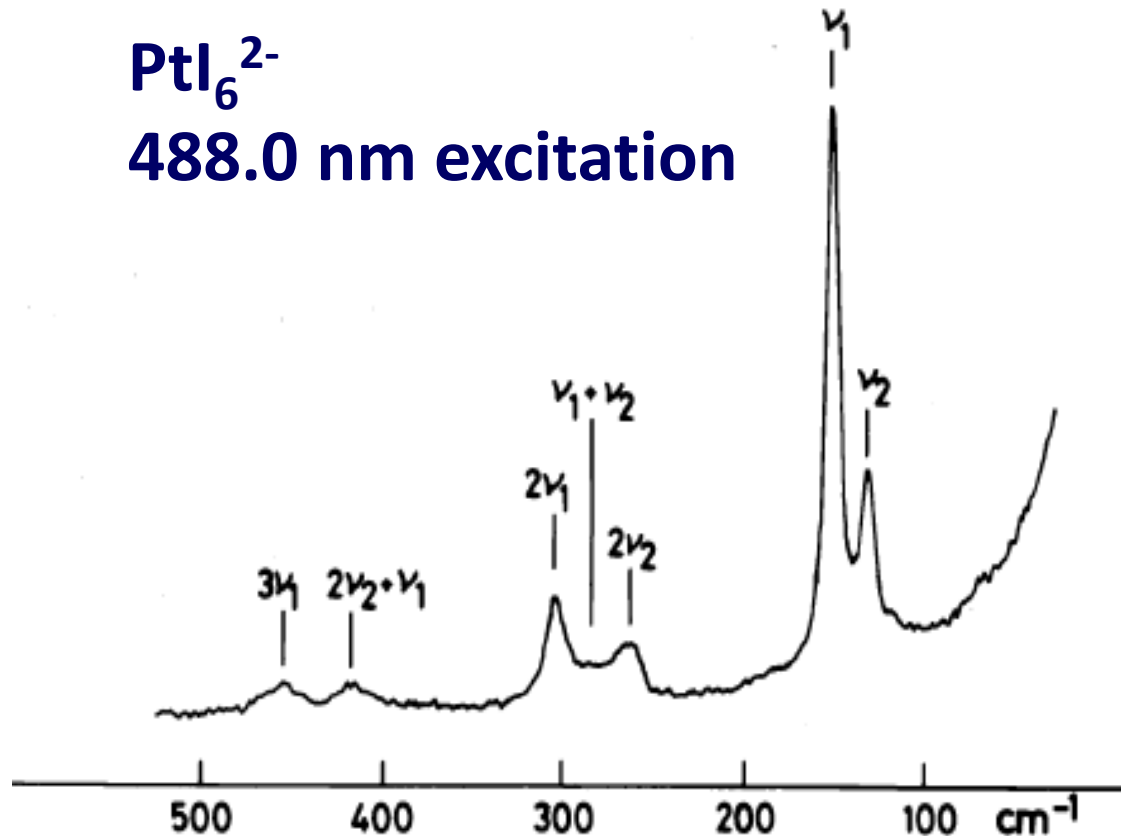
$$t_{2g} \sim \begin{pmatrix} 0 & 1 & 0 \\ 1 & 0 & 0 \\ 0 & 0 & 0 \end{pmatrix}$$

$\rho=0.75$

# Resonance Raman spectrum of $\text{PtI}_6^{2-}$

$\text{PtI}_6^{2-}$

488.0 nm excitation

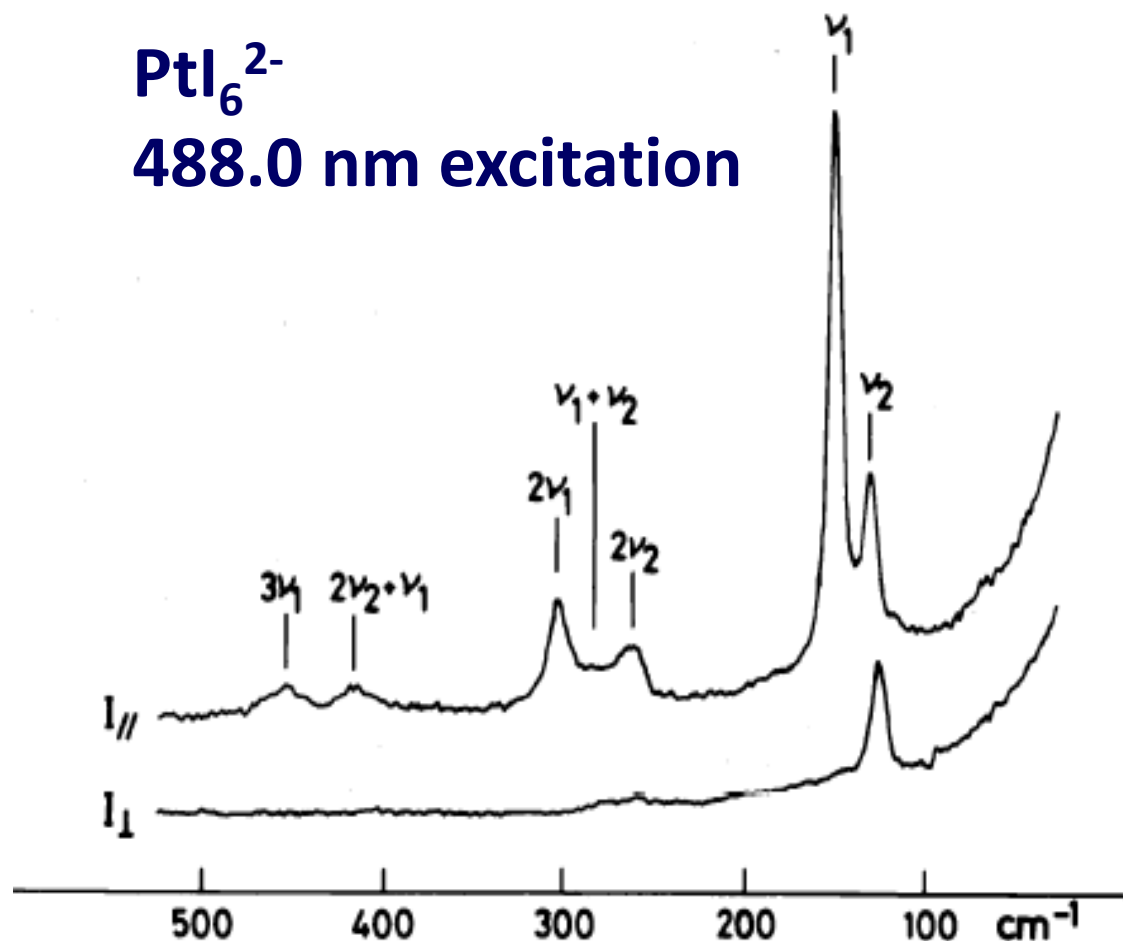


Totally symmetric mode ( $\nu_1, 2\nu_1, 3\nu_1$ )  $\rightarrow$  **A term**

Non-totally symmetric mode ( $\nu_2, 2\nu_2, \nu_1 + \nu_2, 2\nu_1 + \nu_2$ )  $\rightarrow$  **B term**

# Polarized Resonance Raman Spectra of $\text{PtI}_6^{2-}$

$\text{PtI}_6^{2-}$   
488.0 nm excitation

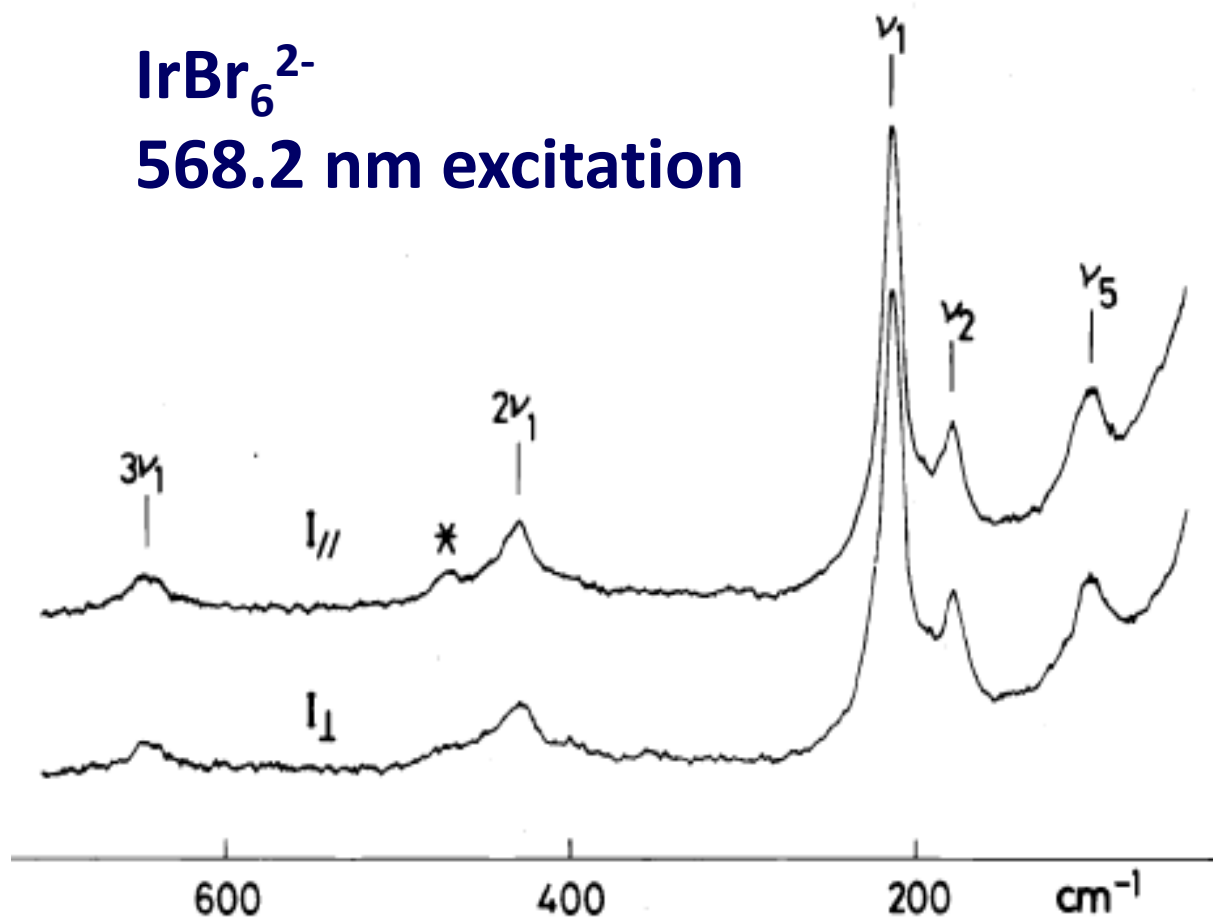


$\nu_1, 2\nu_1, 3\nu_1$  bands  $\rho=0$  ;  $\nu_2$  band  $\rho=0.75$

H. Hamaguchi, *J. Chem. Phys.*, **69**, 569-578 (1978).

# Polarized Resonance Raman Spectra of $\text{IrBr}_6^{2-}$

$\text{IrBr}_6^{2-}$   
568.2 nm excitation



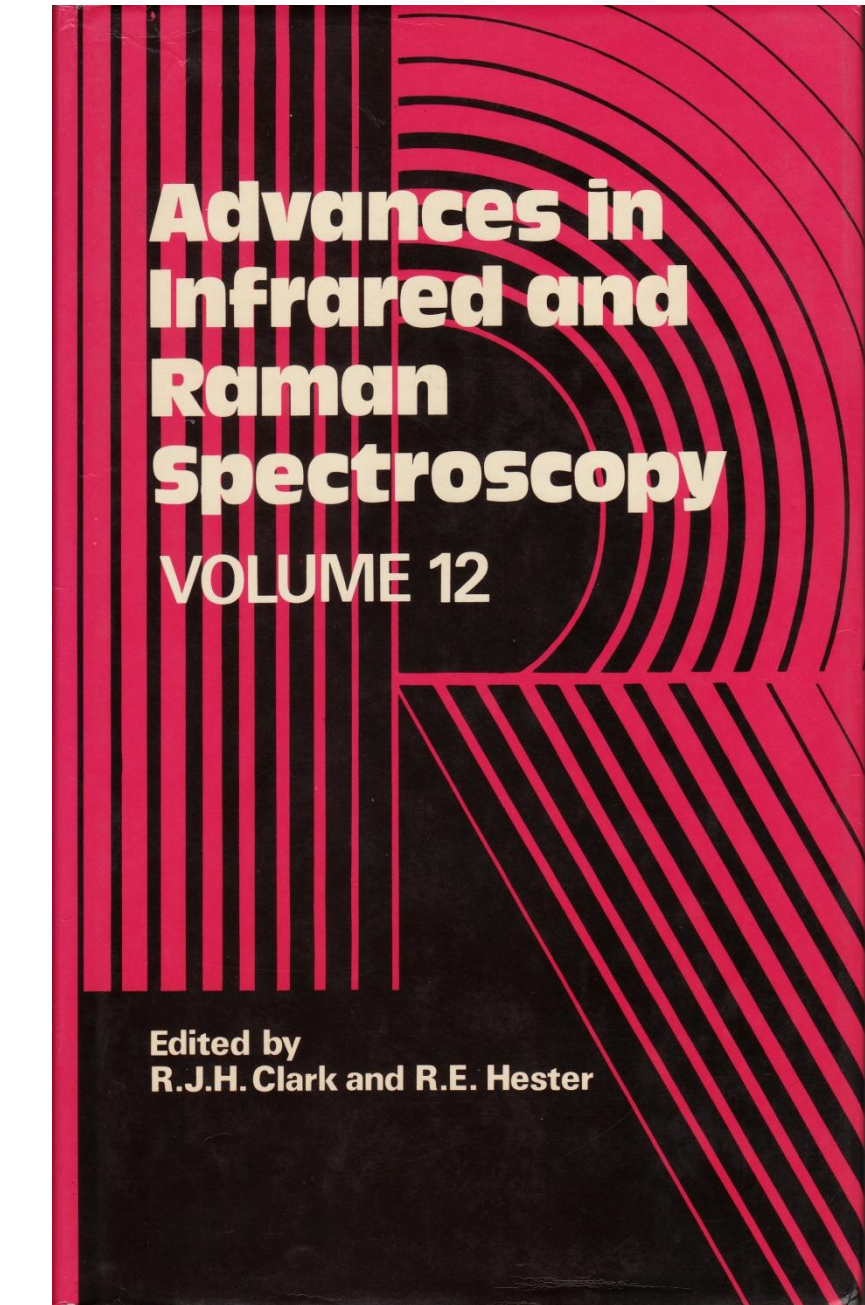
$\rho=1$  for all bands; **forgot to rotate the analyzer?**

# Raman Scattering Tensors and Depolarization ratio of the Totally Symmetric Mode of the $\text{IrBr}_6^{2-}$ Ion

$$\begin{array}{l}
 |g(\alpha)\rangle \longrightarrow |g(\alpha)\rangle \quad \begin{pmatrix} 1 & i & 0 \\ -i & 1 & 0 \\ 0 & 0 & 1 \end{pmatrix} \quad G_0=3, G_a=2, G_s=0 \\
 |g(\alpha)\rangle \longrightarrow |g(\beta)\rangle \quad \begin{pmatrix} 0 & 0 & -i \\ 0 & 0 & -1 \\ i & 1 & 0 \end{pmatrix} \quad G_0=0, G_a=4, G_s=0 \\
 |g(\beta)\rangle \longrightarrow |g(\alpha)\rangle \quad \begin{pmatrix} 0 & 0 & -i \\ 0 & 0 & 1 \\ i & -1 & 0 \end{pmatrix} \quad G_0=0, G_a=4, G_s=0 \\
 |g(\beta)\rangle \longrightarrow |g(\beta)\rangle \quad \begin{pmatrix} 1 & -i & 0 \\ i & 1 & 0 \\ 0 & 0 & 1 \end{pmatrix} \quad G_0=3, G_a=2, G_s=0
 \end{array}$$

$$G_0=6, G_a=12, G_s=0 \quad \rho=(3G_s+5G_a)/(10G_0+4G_s)=1$$





# Advances in Infrared and Raman Spectroscopy VOLUME 12

Edited by  
R.J.H. Clark and R.E. Hester

(1985)

## Chapter 6

# THE RESONANCE EFFECT AND DEPOLARIZATION IN VIBRATIONAL RAMAN SCATTERING

Hiro-o Hamaguchi

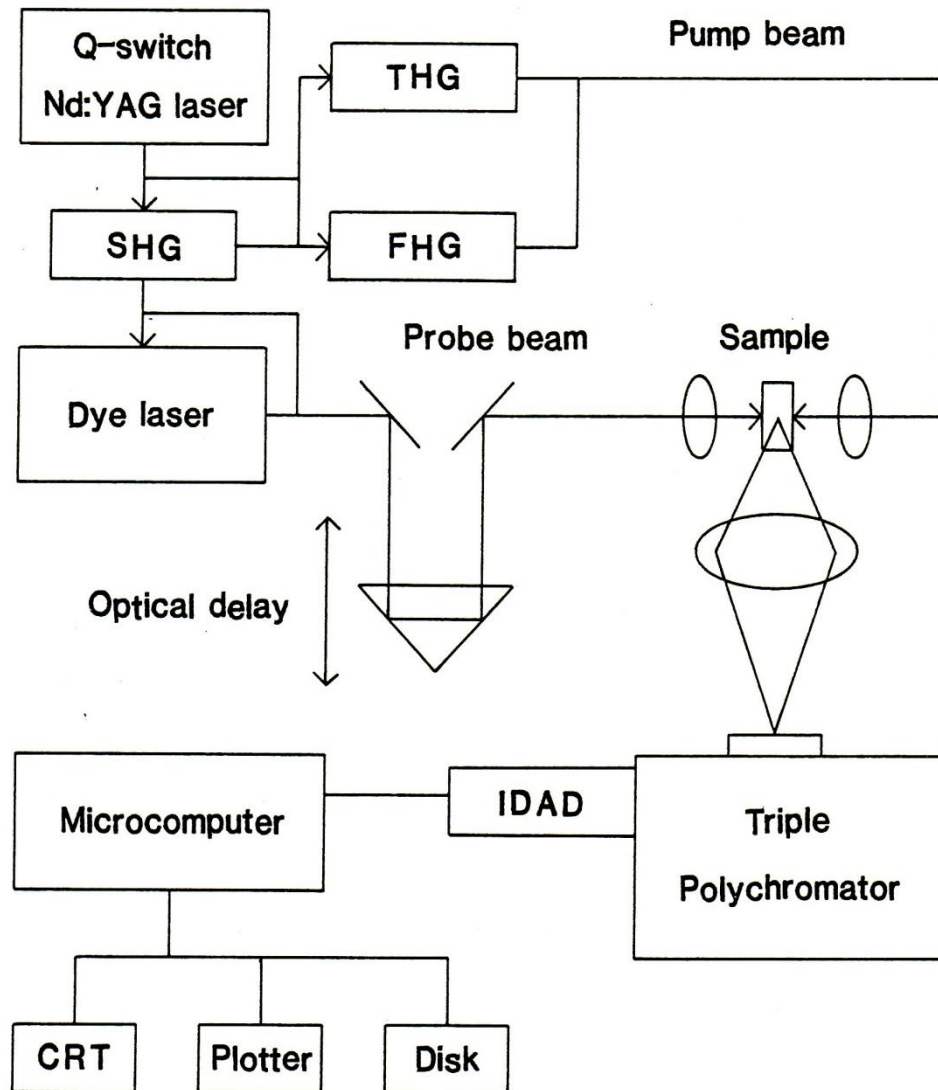
*Department of Chemistry, Faculty of Science, The University of Tokyo, Bunkyo-ku, Tokyo 113, Japan*

## 1 INTRODUCTION

Vibrational Raman scattering is essentially a vibronic process which involves the initial, intermediate, and final vibronic states. Under certain conditions, however, it can be regarded as a purely vibrational process similar to infrared absorption. This was first shown by Placzek,<sup>(1)</sup> who proved that if (i) the ground electronic state is nondegenerate and (ii) the excitation is off-resonant, the vibrational Raman intensities are approximately given by the vibrational matrix elements at the electronic polarizability (the Placzek polarizability theory). Conditions (i) and (ii) are well satisfied for off-resonant vibrational Raman scattering from molecules in their nondegenerate ground electronic states and the polarizability theory has been extensively used by vibrational spectroscopists, who found it convenient to treat vibrational Raman scattering with an exact parallelism to infrared absorption; the intensity of infrared absorption is related to the vibrational-coordinate dependence of the dipole moment, while the vibrational Raman intensity is correlated with that of the polarizability. The Placzek polarizability theory has another advantage in that it is in harmony with the classical picture of vibrational Raman scattering, in which the oscillating dipole moment induced by the electric field of the incident light is modulated by the vibrational motions, resulting in scattering with shifted frequencies.

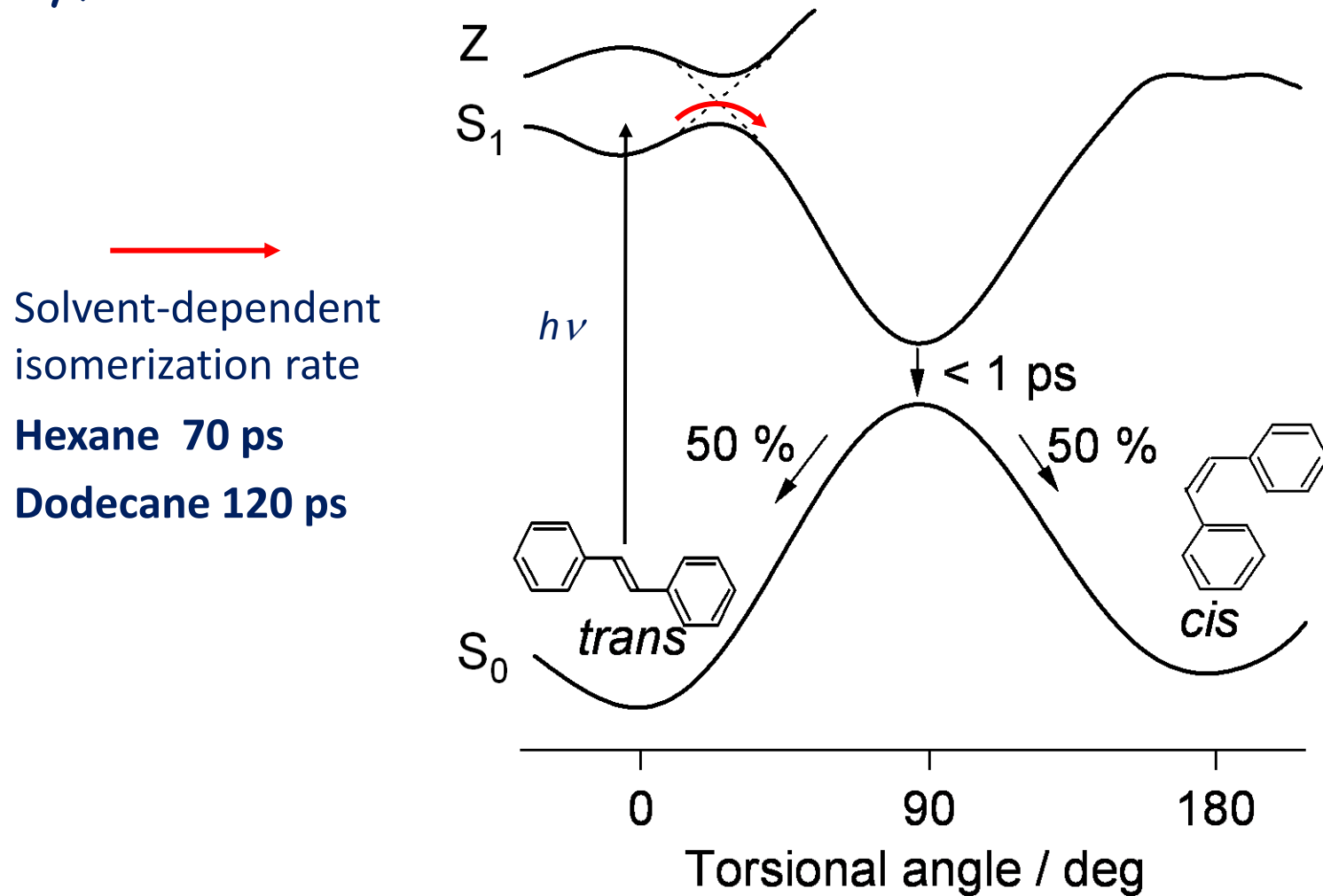
The breakdown of the polarizability theory manifests itself most clearly in the polarization properties of vibrational Raman scattering. As predicted by Placzek,<sup>(1)</sup> the removal of either or both of conditions (i) and (ii) may lead

# Nanosecond Transient Raman Spectrometer (1983)



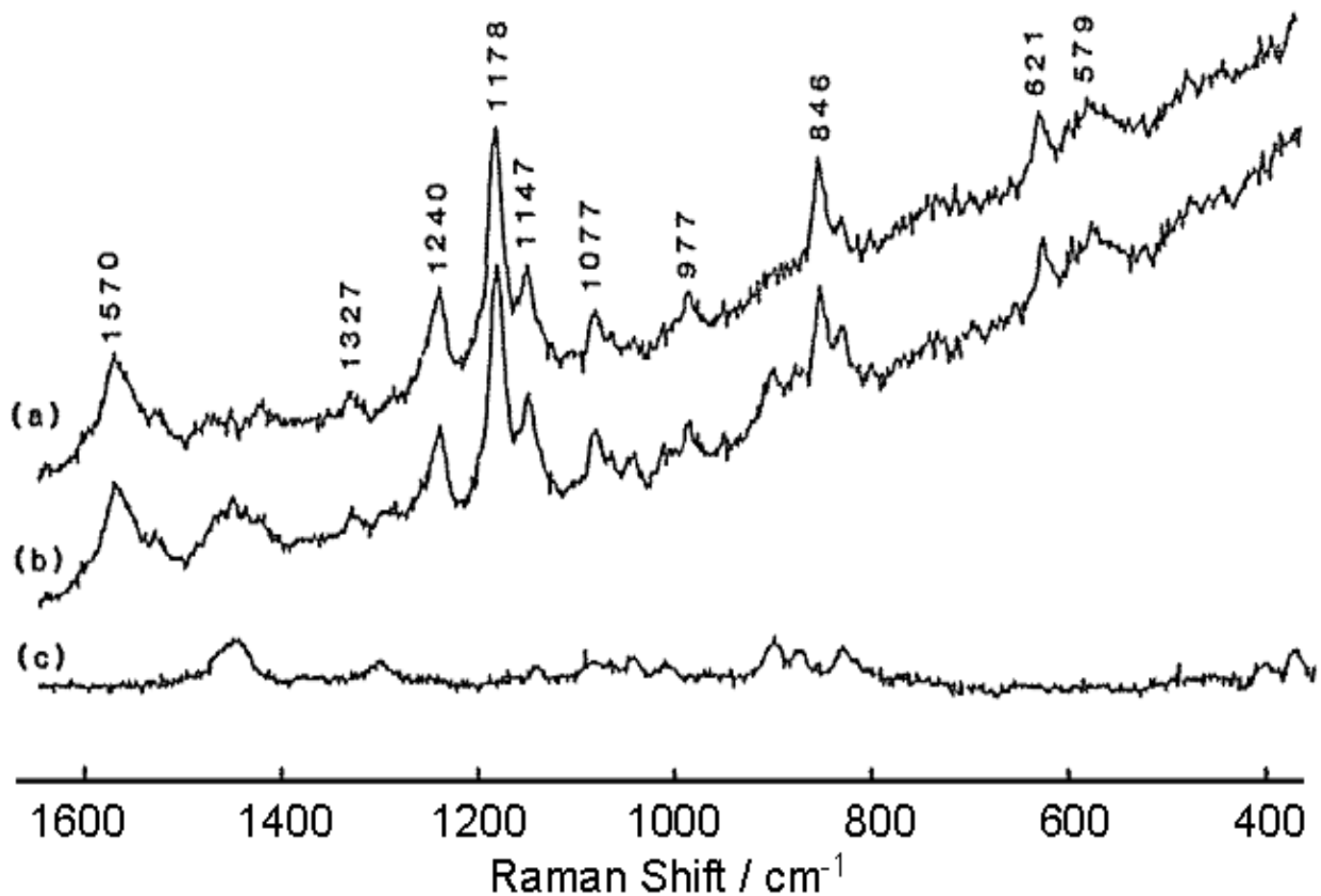
# Photoisomerization of *Trans*-Stilbene

Why, when and how rotation occurs in the excited state?



Probe solvent dependent structure and dynamics of  $S_1$  *trans*-stilbene by time-resolved Raman spectroscopy

# Nanosecond Transient Raman Spectrum of $S_1$ *Trans*-Stilbene (1983)

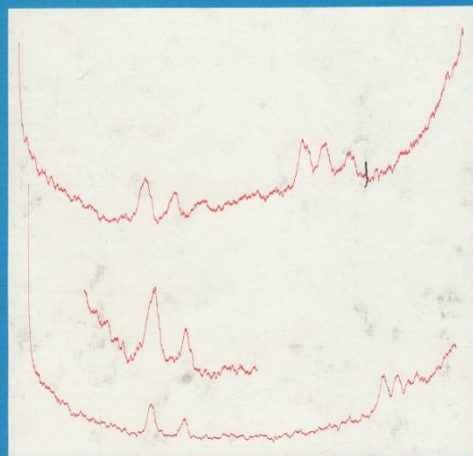


T. L. Gustafson, D. M. Roberts, and D. A. Chernoff, *J. Chem. Phys.* 79, 1559 (1983).  
H. Hamaguchi, C. Kato, M. Tasumi, *Chem. Phys. Lett.*, **100**, 3-7 (1983).

# Three Raman Stilbenists on a Bridge near the Noishvanstein Castle (1985)



# VIBRATIONAL SPECTRA AND STRUCTURE



Volume 16

edited by James R. Durig

(1987)

## Chapter 4

### TRANSIENT AND TIME-RESOLVED RAMAN SPECTROSCOPY OF SHORT-LIVED INTERMEDIATE SPECIES

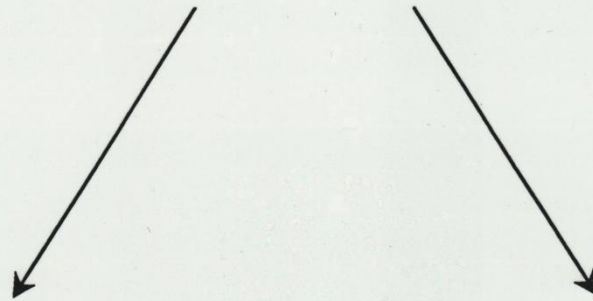
Hiro-o Hamaguchi

Department of Chemistry  
Faculty of Science  
The University of Tokyo  
Tokyo, Japan

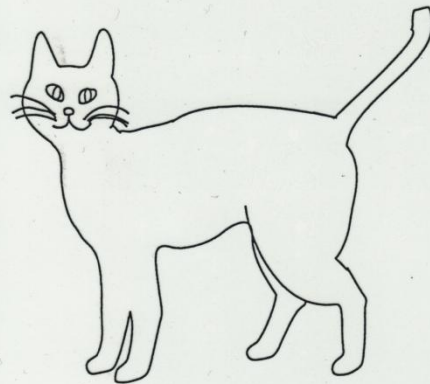
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# KAST

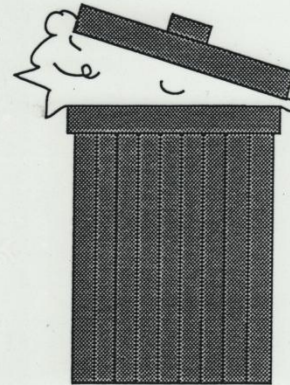
( Kanagawa Academy of  
Science and Technology )

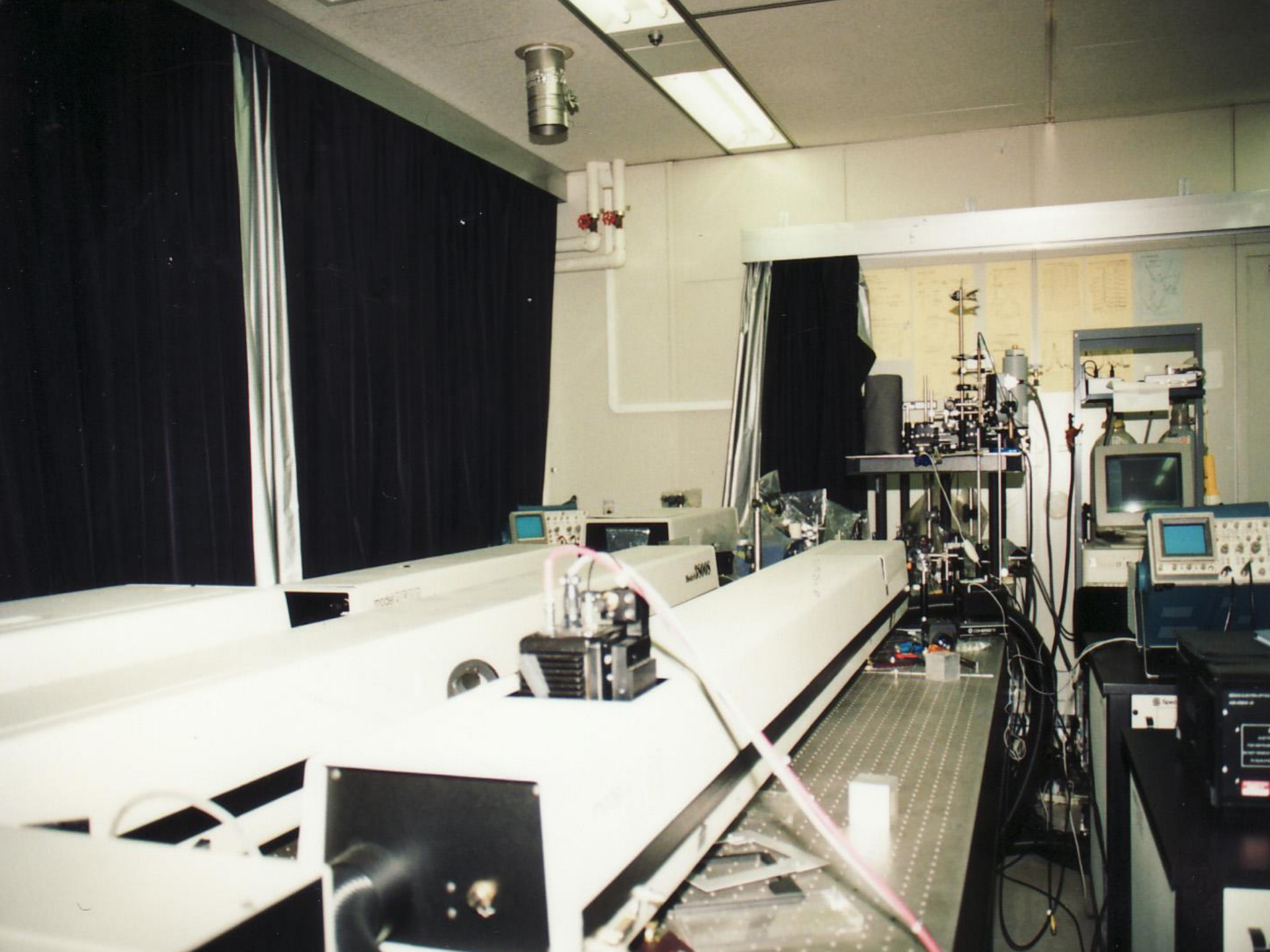


KAT

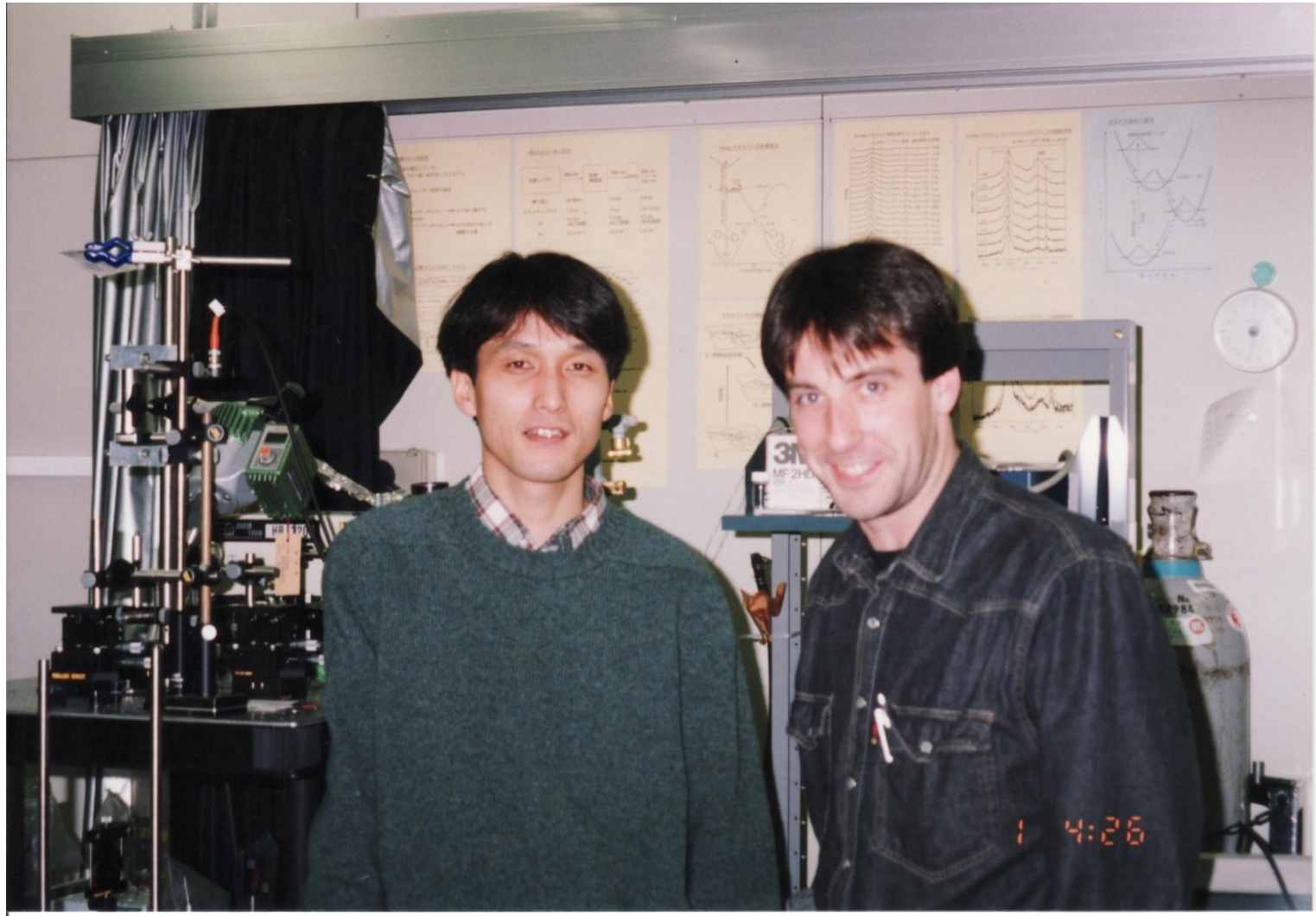


KAS





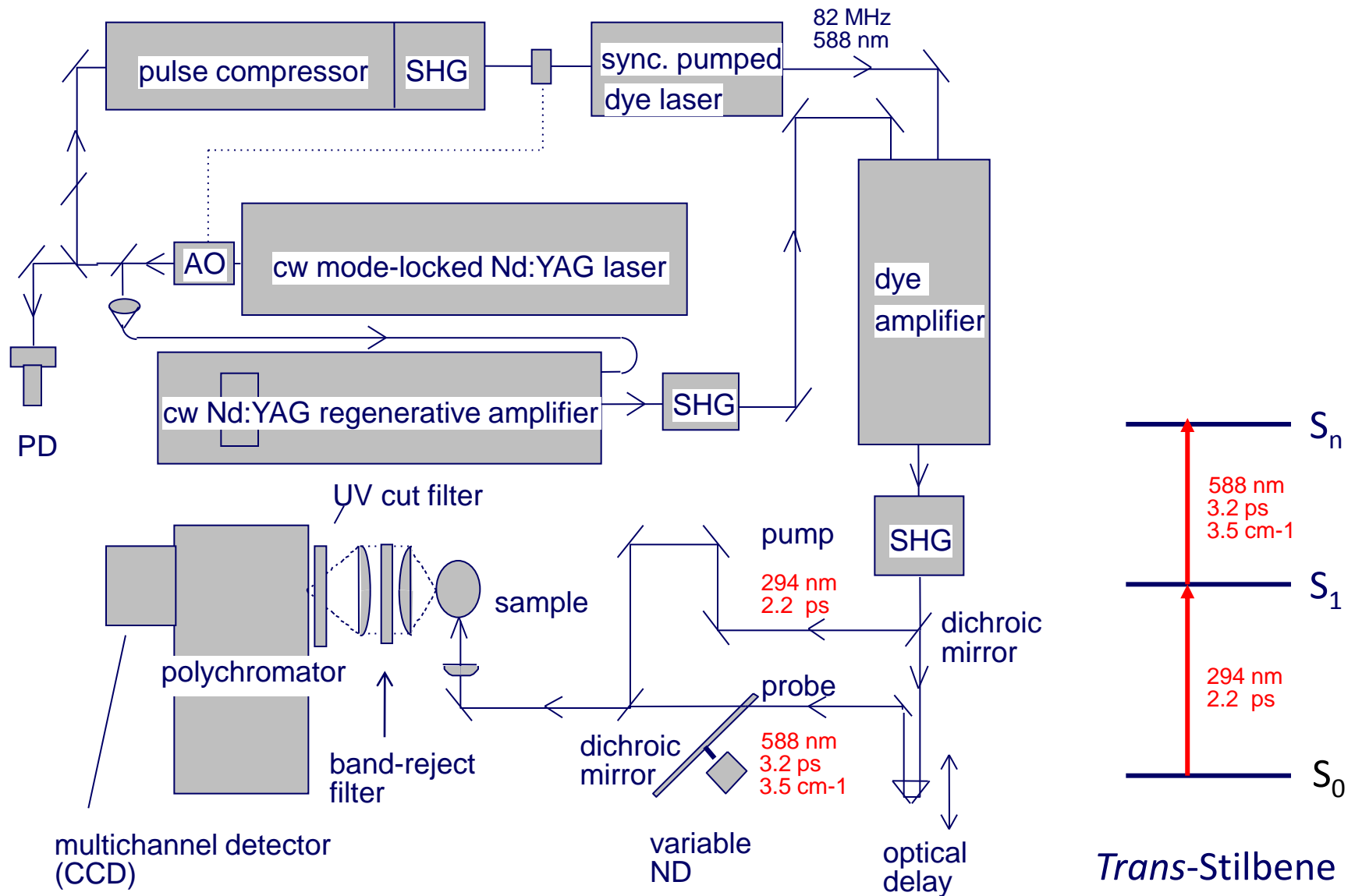




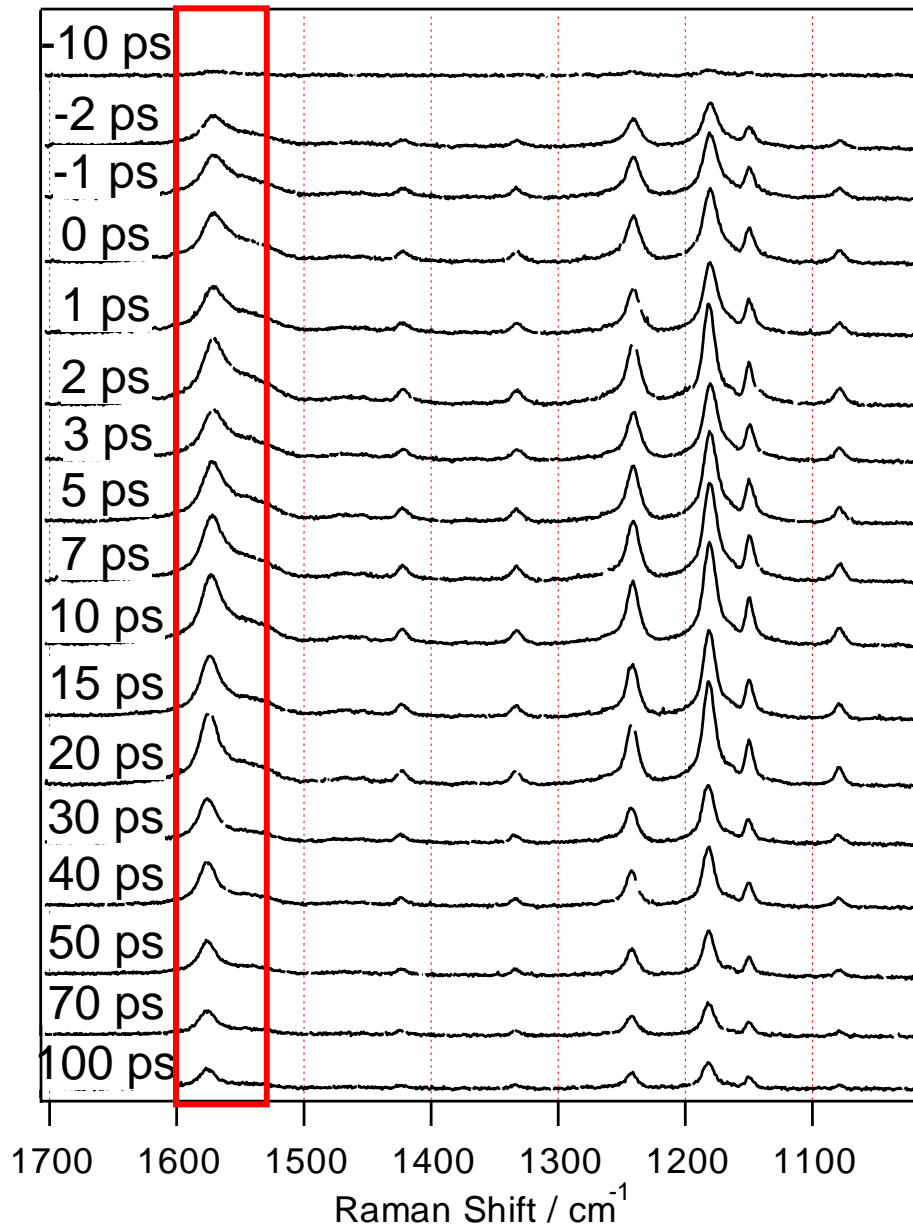
Koichi Iwata

Volker Deckert

# Picosecond Time-resolved Raman Spectrometer



# $S_1$ *trans*-Stilbene in $\text{CHCl}_3$



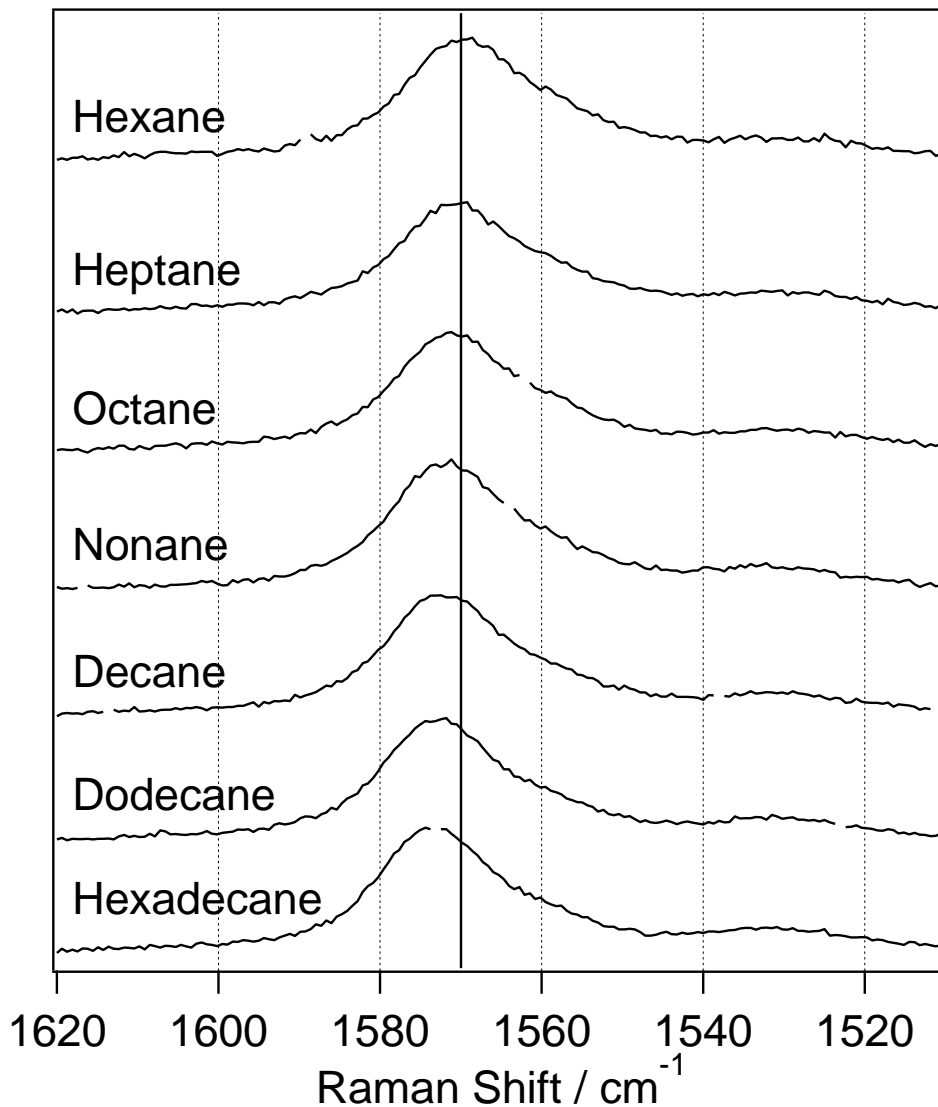
Pump 294 nm  
Probe 588 nm (0.1 mW)

C=C stretch vibration

1560  $\text{cm}^{-1}$ : double bond

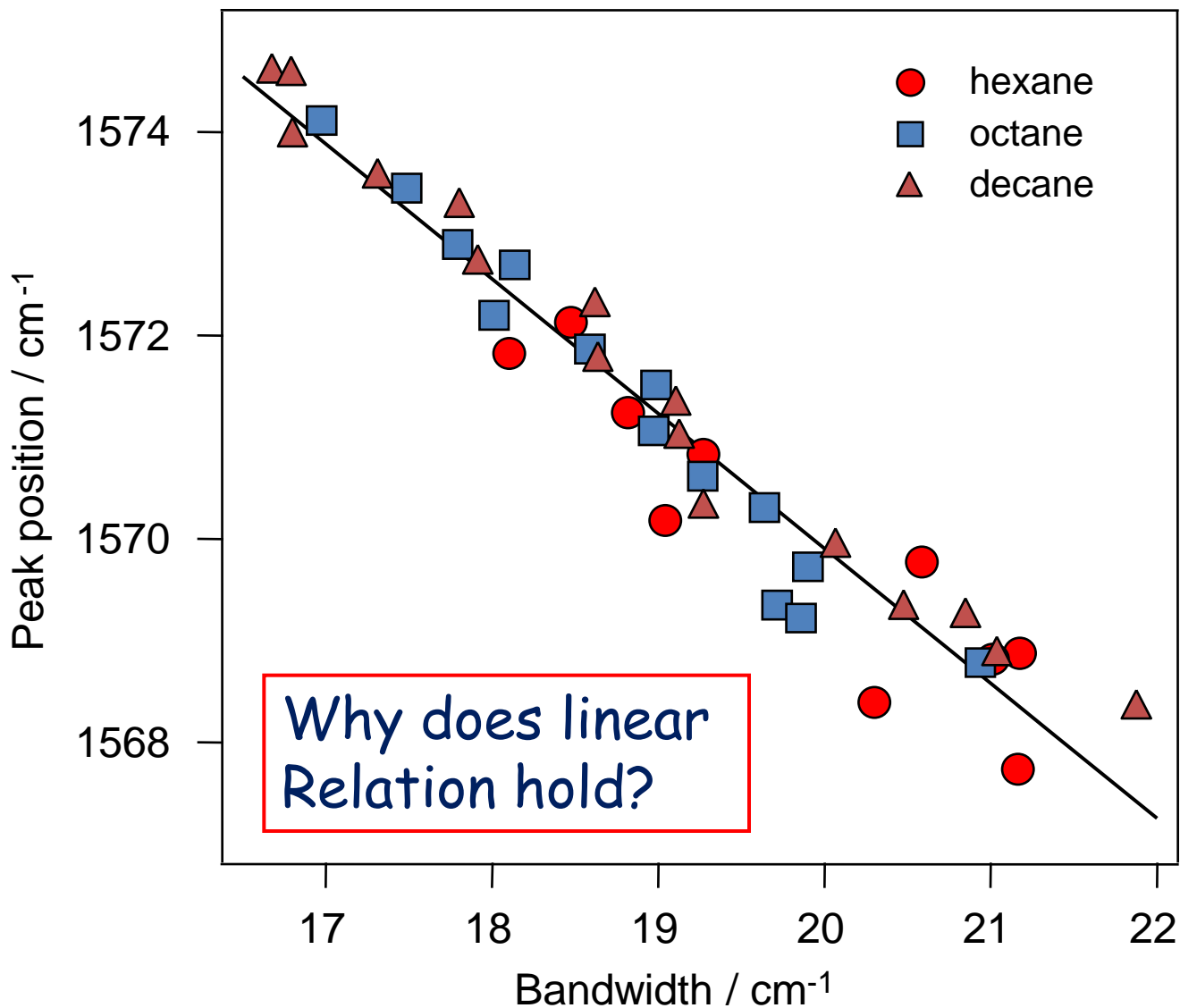
Why rotation occurs  
around a double bond ?

# The C=C Stretch Raman Band of $S_1$ *trans*-Stilbene in Alkanes



The peak position shifts to higher wavenumbers and the band width decreases on going from hexane to hexadecane. Why?

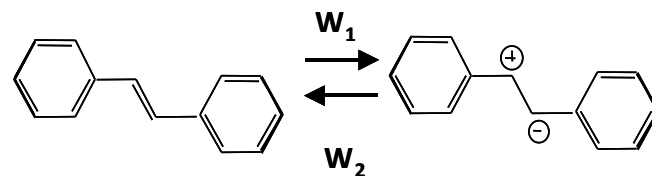
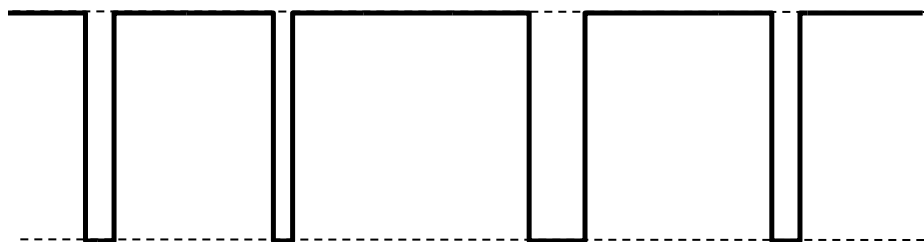
# Peak Position vs Band Width in Alkane Solvents at Different Temperatures



# Dynamic Frequency Exchange Model and Vibrational Bandshapes

Hamaguchi *Mol. Phys.* **89**, 463 (1997).

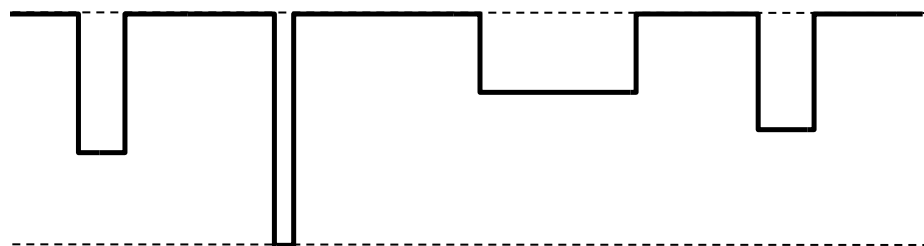
(a) Two Frequency Exchange Model



$$\Delta\Omega = W_1\tau/(1+\tau^2) \quad \Delta\Gamma = W_1\tau^2/(1+\tau^2)$$

$$\Delta\Gamma / \Delta\Omega = \tau$$

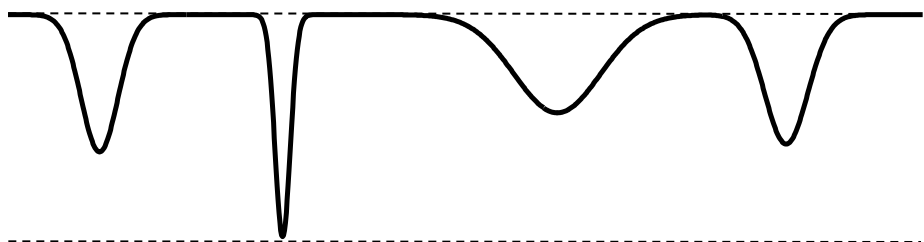
(b) Many Frequency Exchange Model



$$\Delta\Omega = W_1 \sum_{\kappa=2}^n (W_{1\kappa} / W_1) \tau_{\kappa} / (1 + \tau_{\kappa}^2),$$

$$\Delta\Gamma = W_1 \sum_{\kappa=2}^n (W_{1\kappa} / W_1) \tau_{\kappa}^2 / (1 + \tau_{\kappa}^2),$$

(c) Continuous Frequency Modulation Model

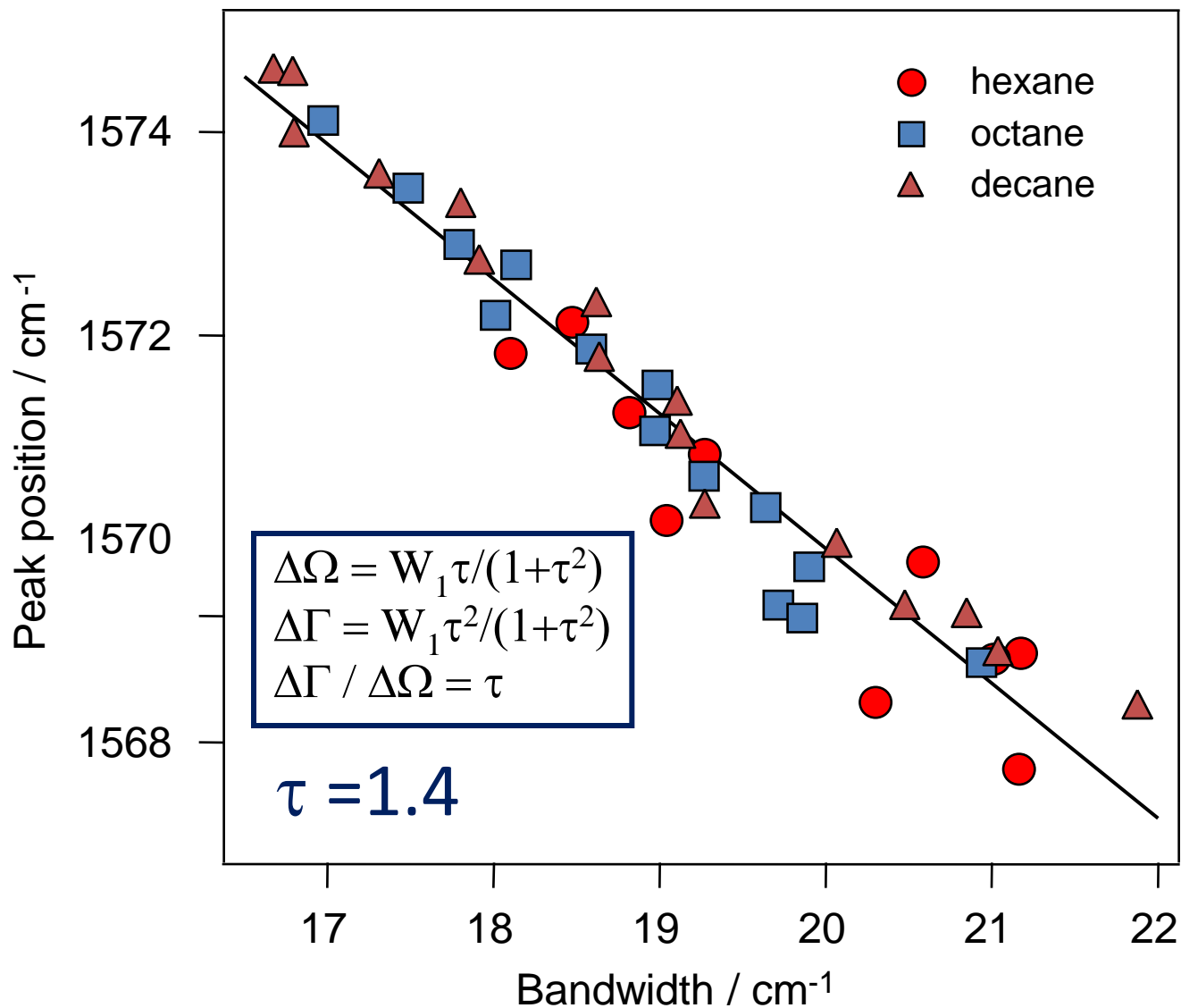


$$\Delta\Omega = \frac{W_1 \int_{-\infty}^0 G(\tau) \tau / (1 + \tau^2) d\tau}{\int_{-\infty}^0 G(\tau) d\tau}$$

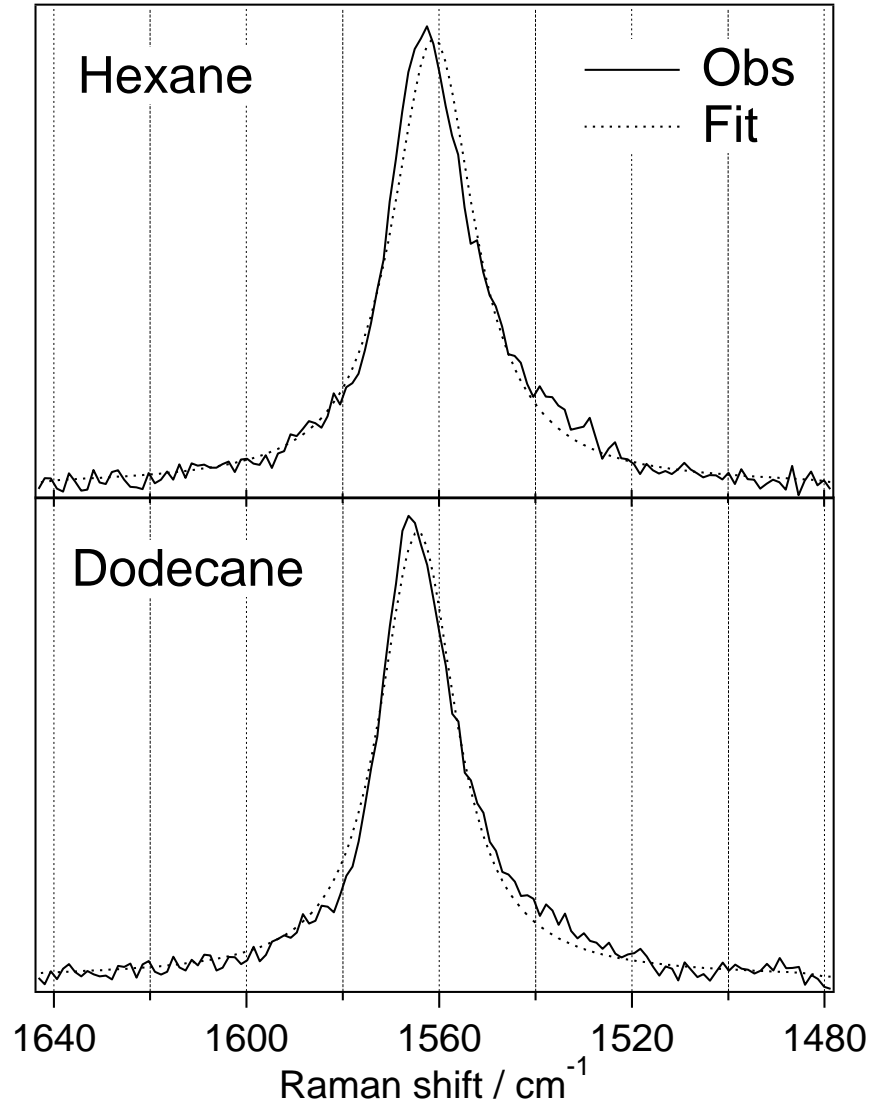
$$\Delta\Gamma = \frac{W_1 \int_{-\infty}^0 G(\tau) \tau^2 / (1 + \tau^2) d\tau}{\int_{-\infty}^0 G(\tau) d\tau}$$

$$\Delta\Gamma / \Delta\Omega = \tau_{1/2}; \quad G(\tau) = \exp(-\ln 2 \tau^2 / \tau_{1/2}^2)$$

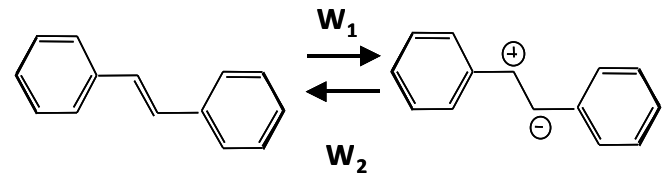
# Peak Position vs Band Width in Alkane Solvents at Different Temperatures



# Fitting of the C=C Stretch Raman Band of *S1 trans*-Stilbene by the Two Frequency Exchange Model



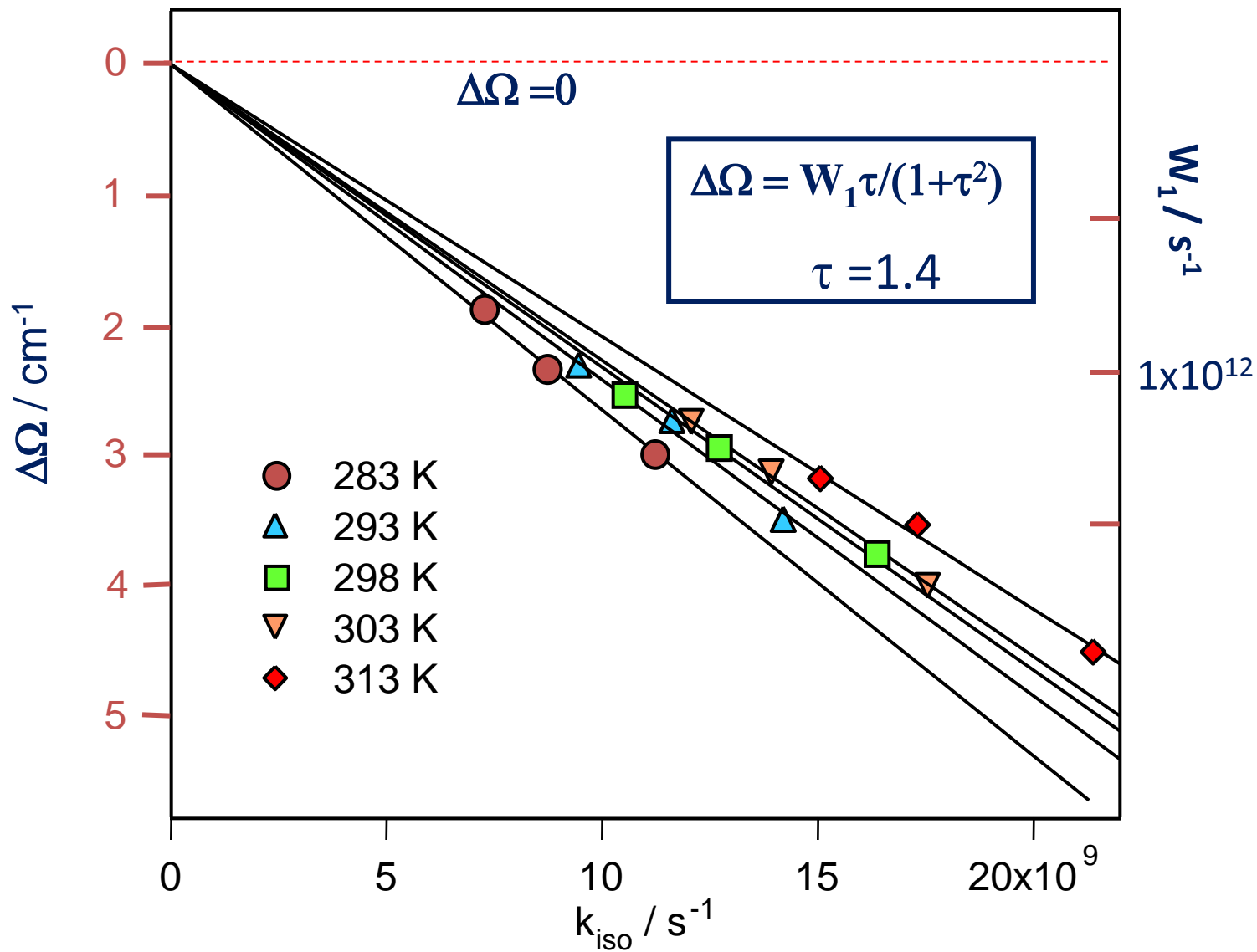
$$W_1 = 2.7 \times 10^{12} \text{ sec}^{-1} \text{ (370 fs)}^{-1}$$



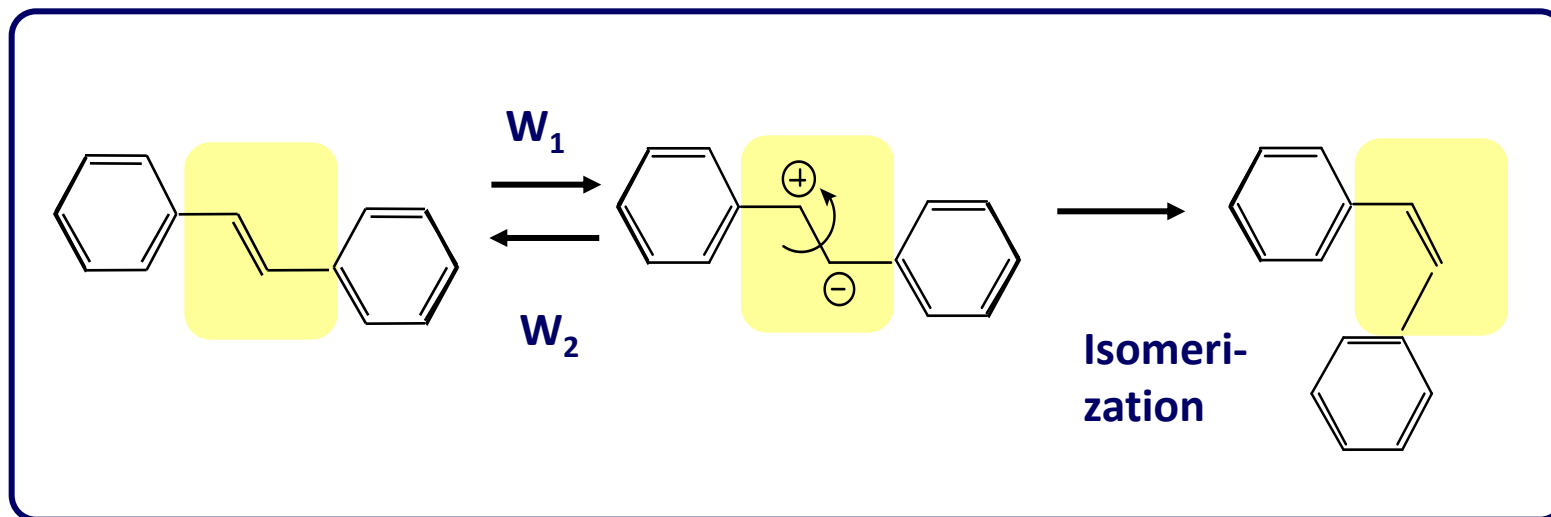
$$W_1 = 1.5 \times 10^{12} \text{ sec}^{-1} \text{ (670 fs)}^{-1}$$



# Isomerization Rate $k_{iso}$ is Proportional to $W_1$



# Dynamic Polarization Model of Isomerization

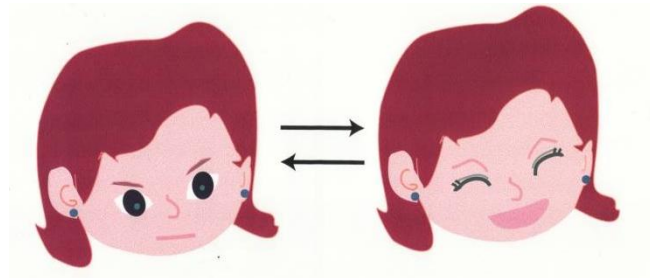
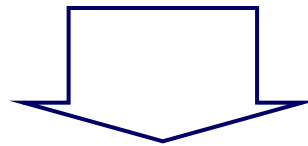
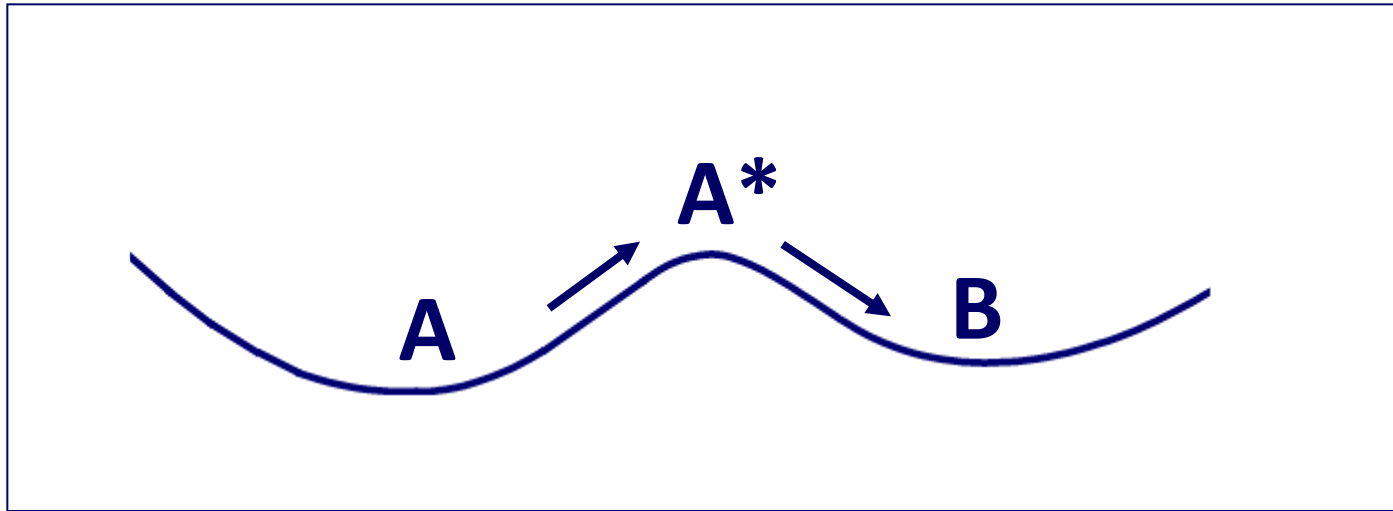


Hamaguchi, Iwata, *CPL* **208**, 465 (1993).

Deckert, Iwata, Hamaguchi, *J. Photochem. Photobiol.* **102**, 35 (1996).

Iwata, Ozawa, Hamaguchi, *JCP* **106**, 3614 (2002).

# How Do Chemical Reactions Proceed?



Cf. Michaelis-Menten kinetics

"How can the events in space and time which take place within the spatial boundary of a living organism be accounted for by physics and chemistry?"

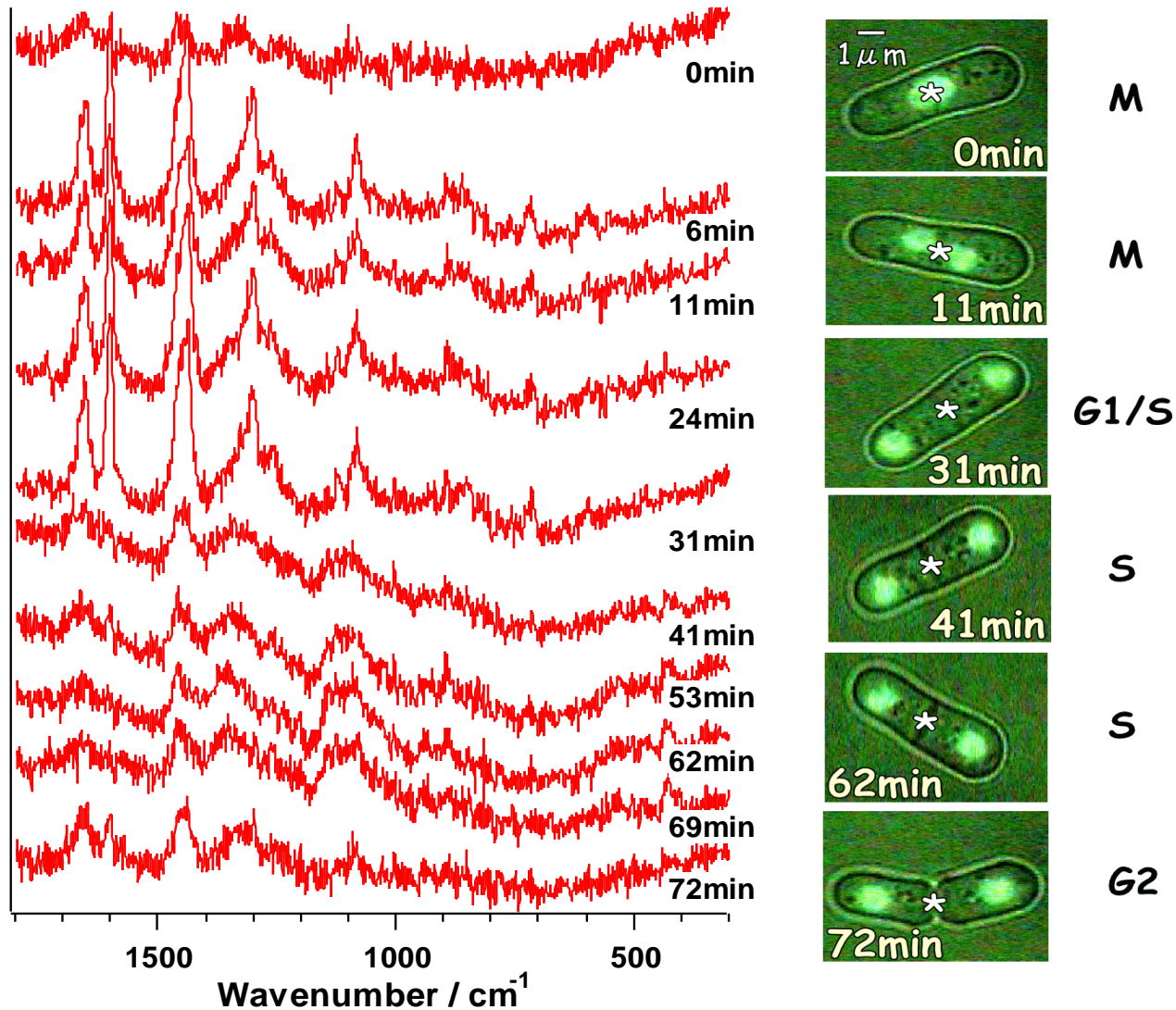
In "What is Life"

Erwin Schrödinger

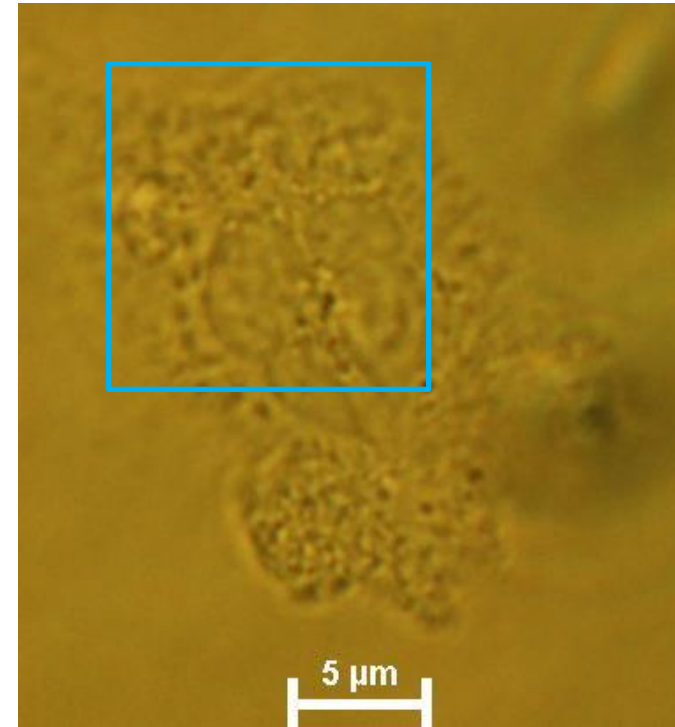
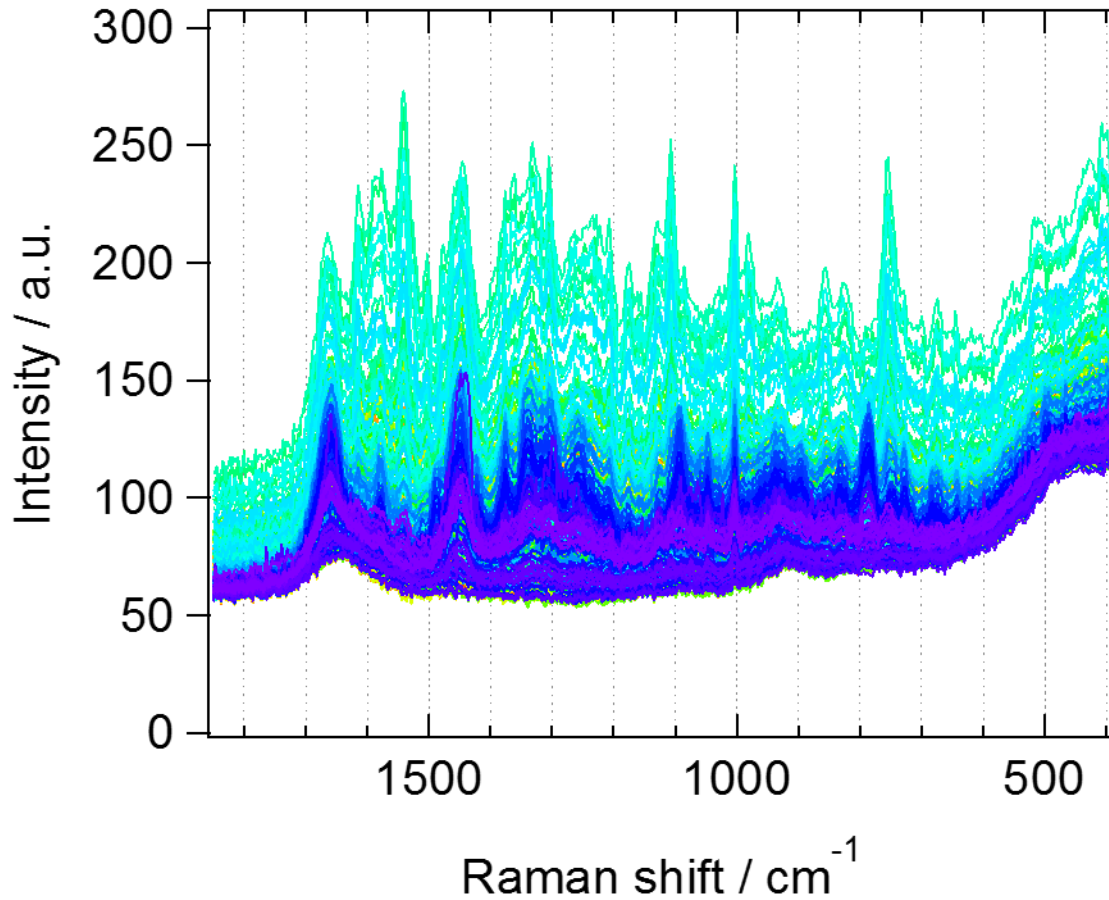


E. Schrödinger  
(1887-1961)

# Raman Spectroscopy of Really Living Cell



# Molecular Component Distribution Imaging (MCDI) of Living Cells with Multivariate Curve Resolution Analysis



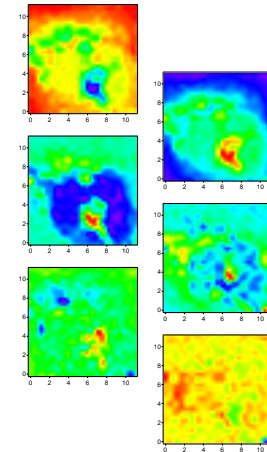
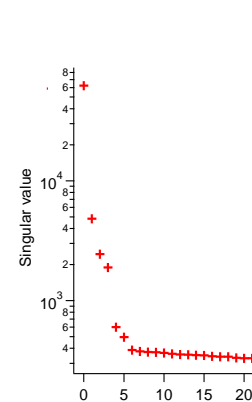
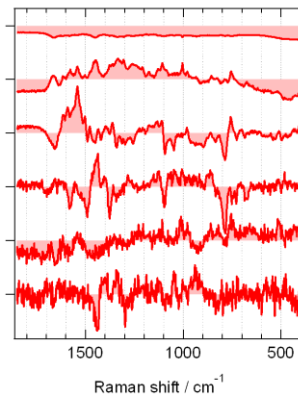
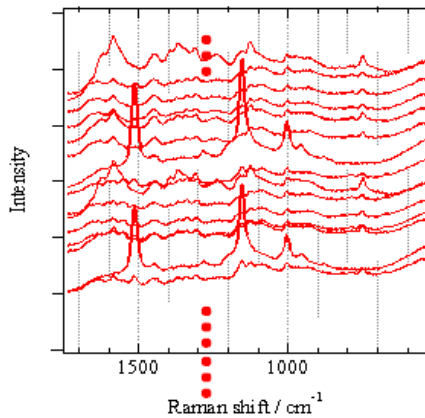
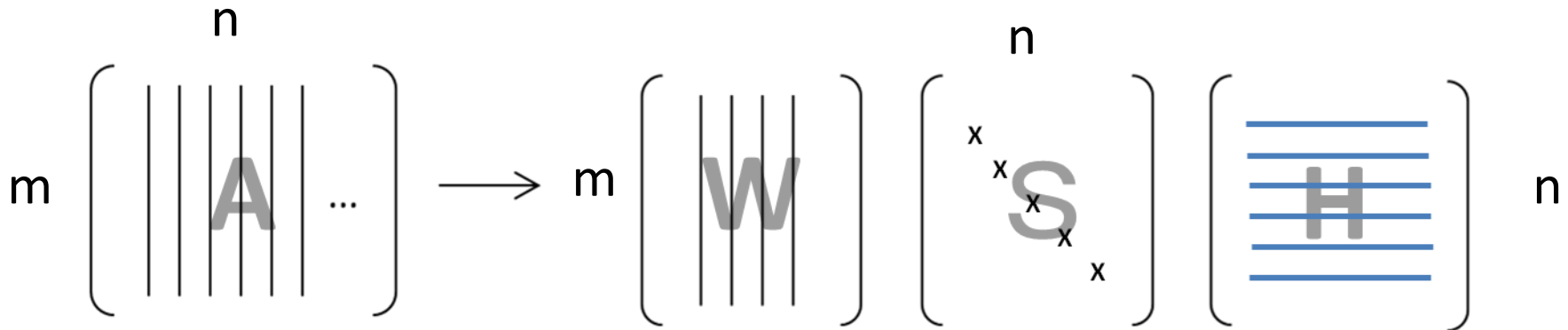
white blood cell

Thorough interpretation of complicated spectra is very difficult.

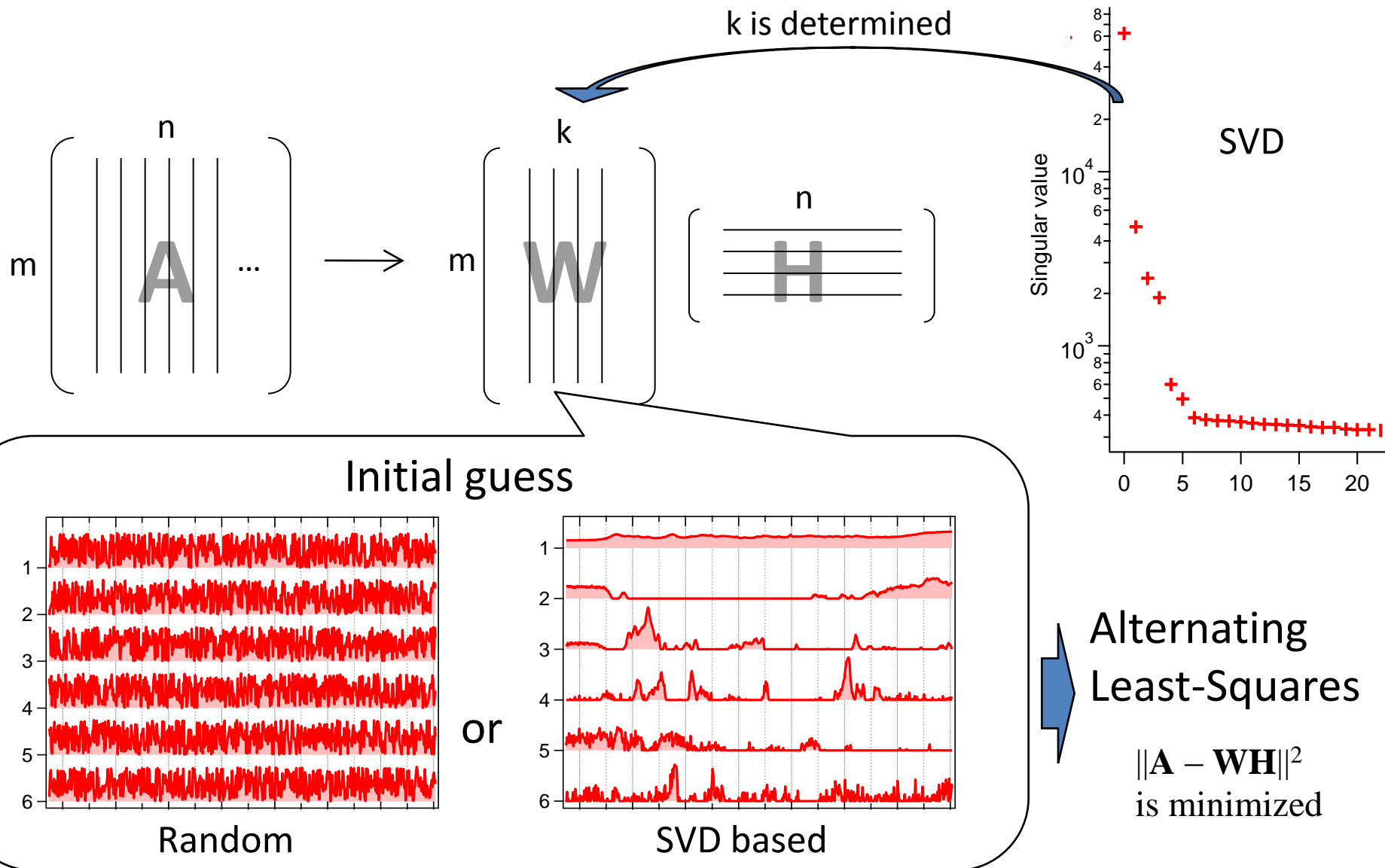
# Multivariate Analysis

## Matrix factorization by Singular Value Decomposition (SVD)

$$A \approx WSH$$



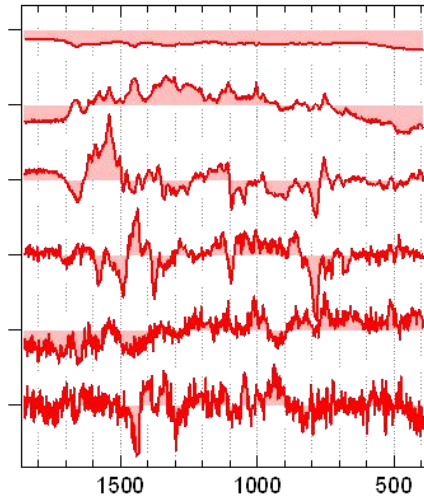
# Multivariate Curve Resolution (MCR)





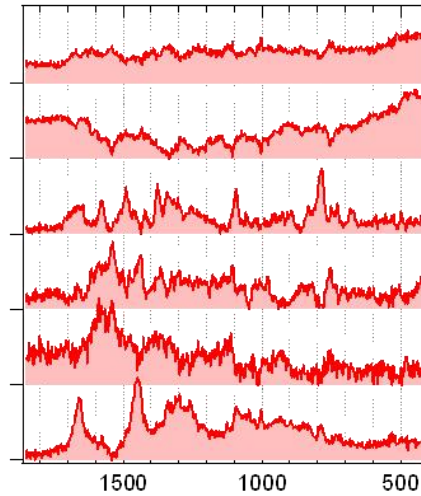
# Multivariate Analyses: Comparison

SVD



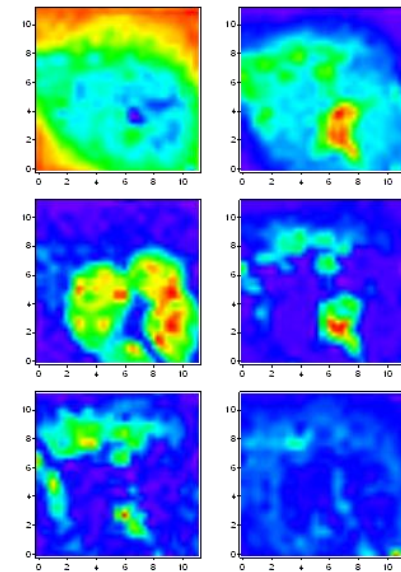
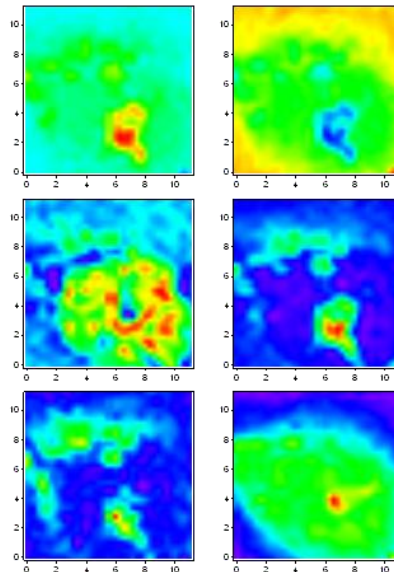
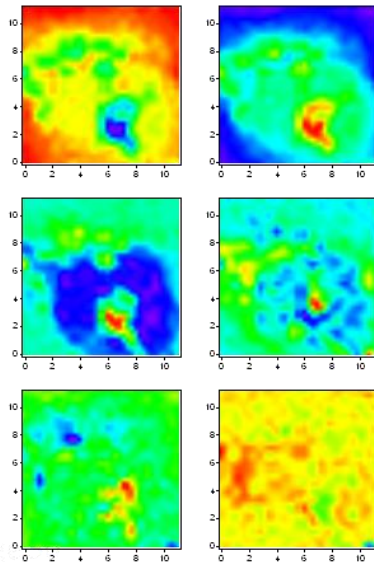
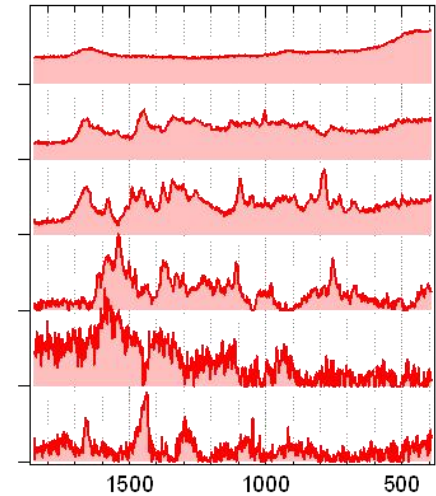
MCR

$W, H \geq 0, L_1 = 0$



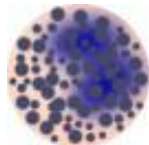




MCR

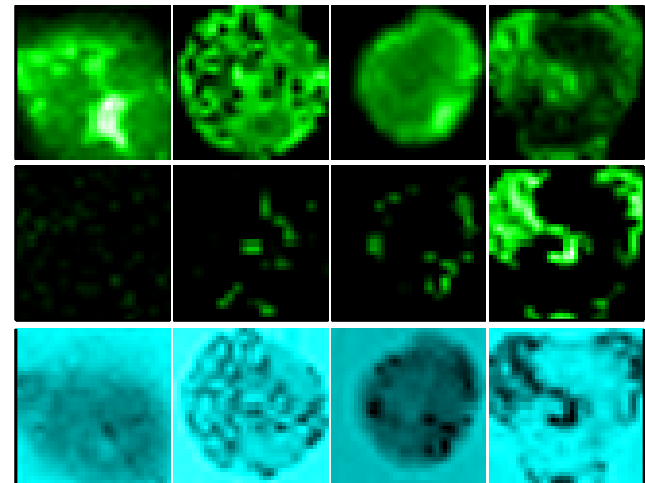
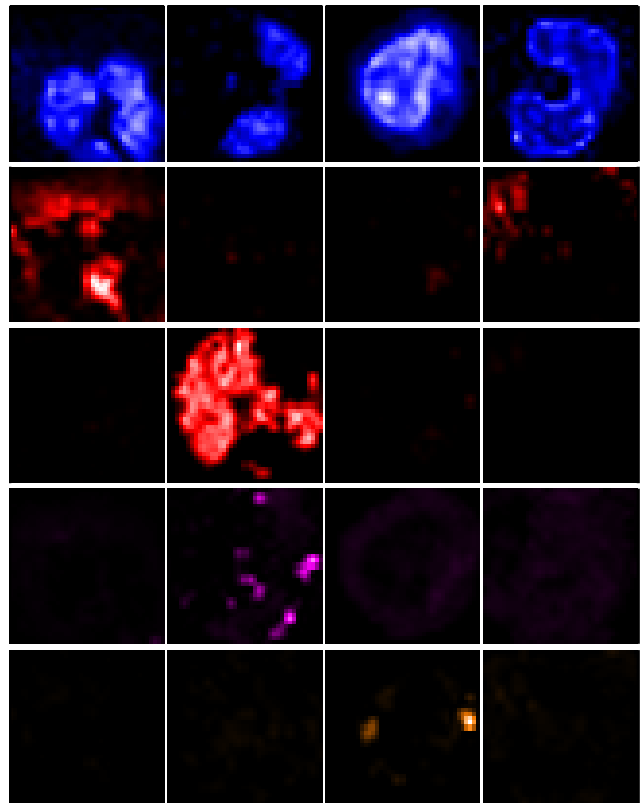
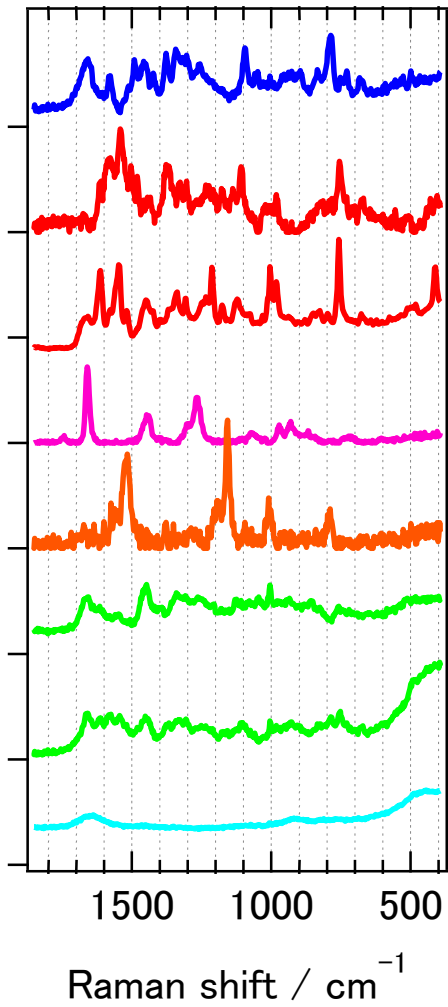
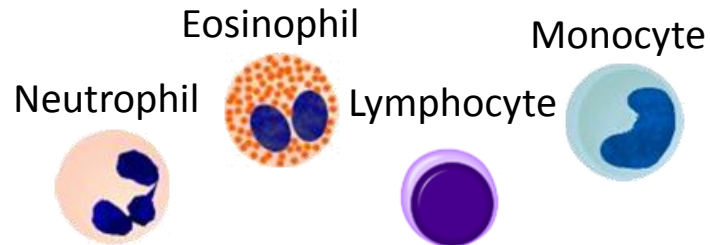
$W, H \geq 0, L_1 = 0.001$



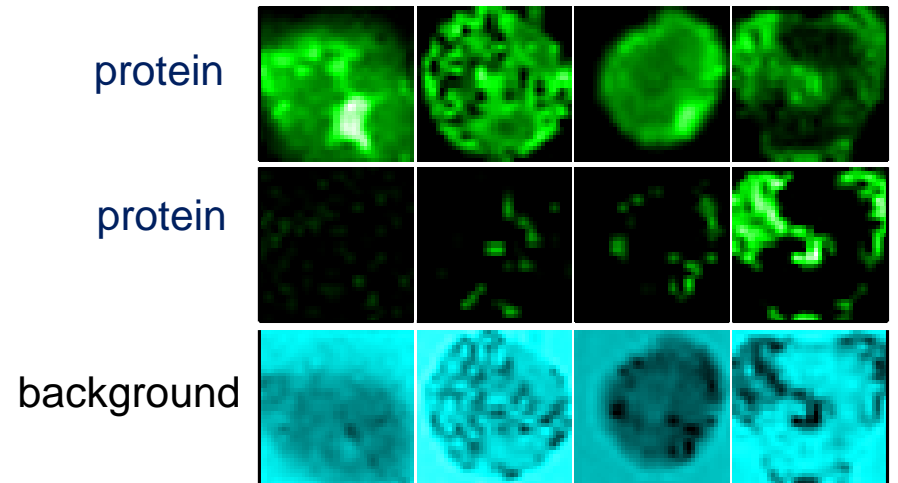
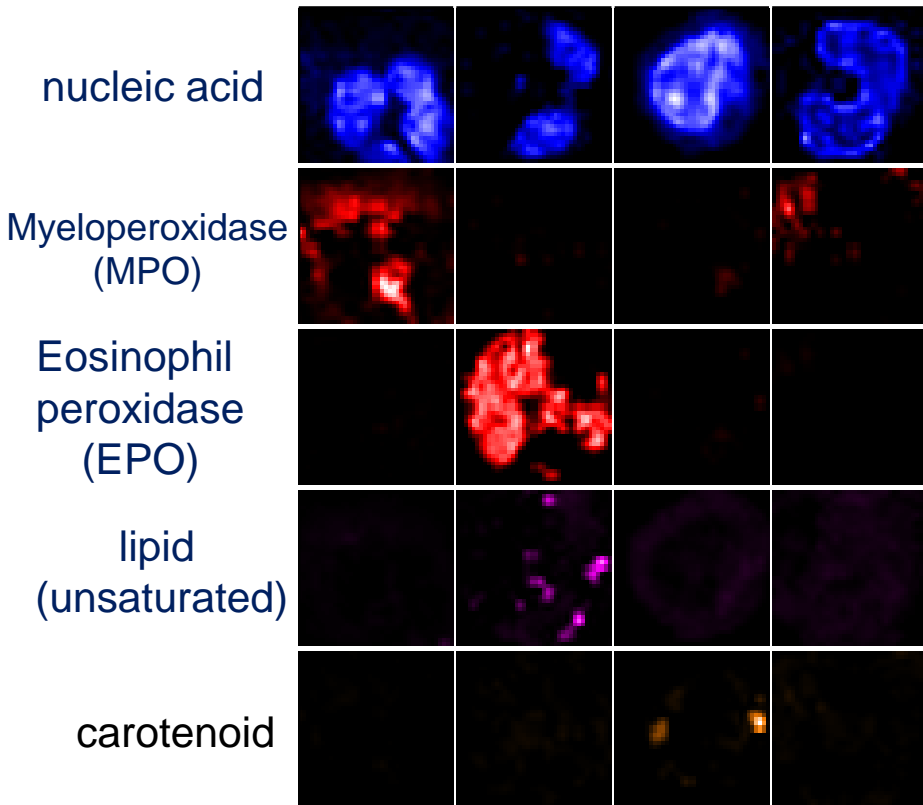
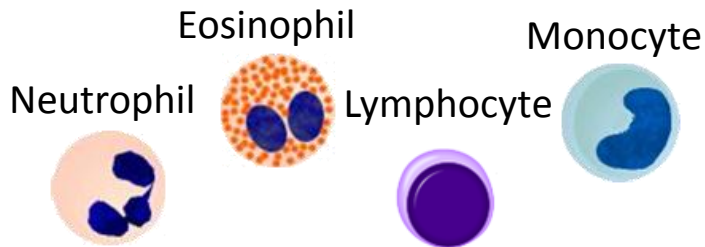
# White Blood Cells

Neutrophil		50-70 %	10-12 $\mu\text{m}$	Phagocytosis - bacteria, fungi
Eosinophil		2-5 %	10-12 $\mu\text{m}$	Combating parasites Modulate allergic inflammatory responses
Basophil		< 1 %	12-15 $\mu\text{m}$	release histamine for inflammatory responses
Lymphocyte		20-40 %	7-8 $\mu\text{m}$	B cells T cells NK cells
Monocyte		3-6 %	14-17 $\mu\text{m}$	Differentiate into tissue resident macrophages

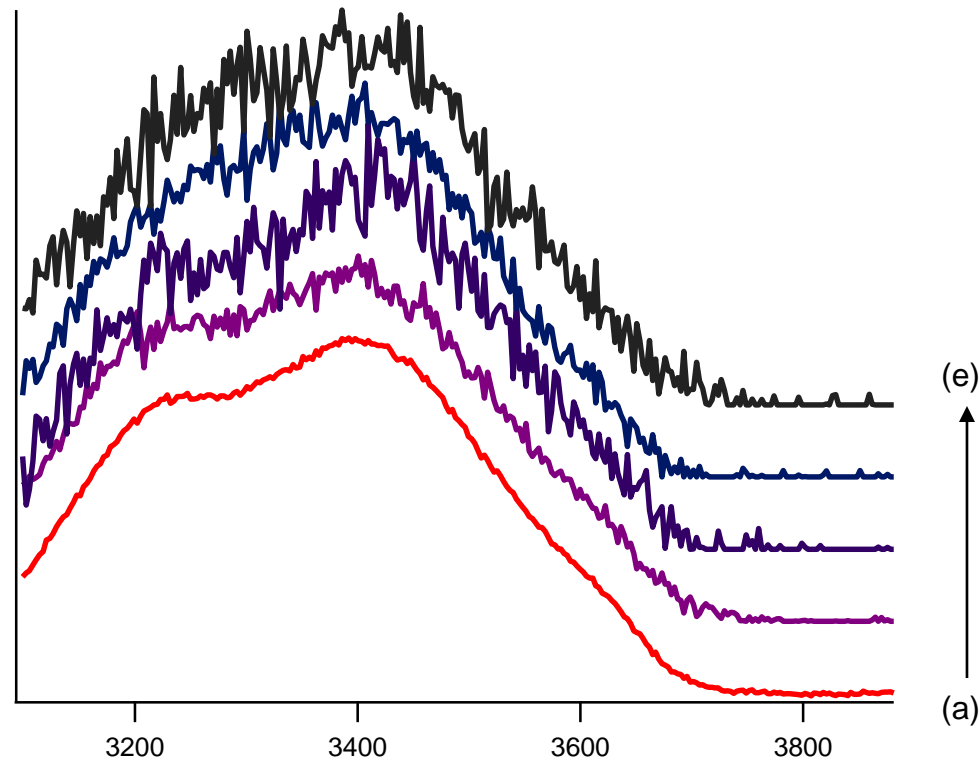
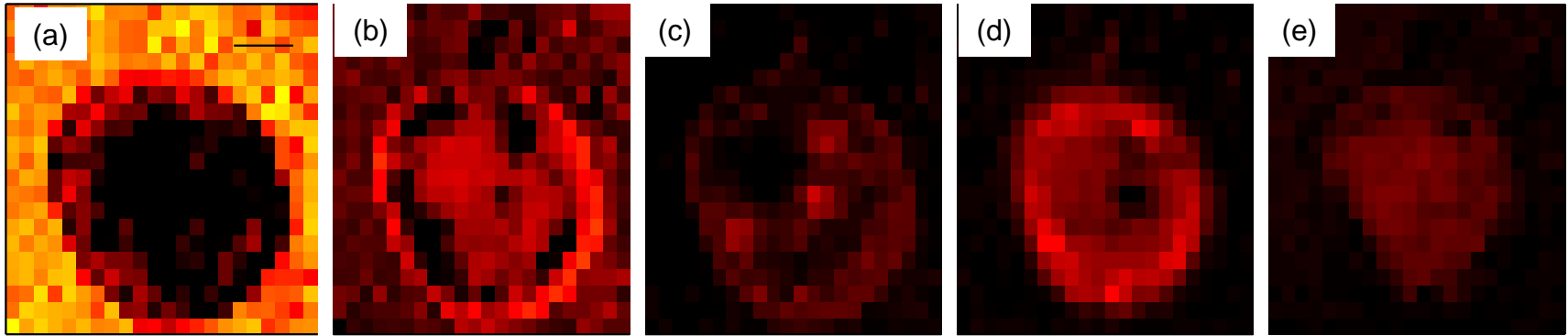
# MCR Analysis of White Blood Cells



# MCDI of White Blood Cells

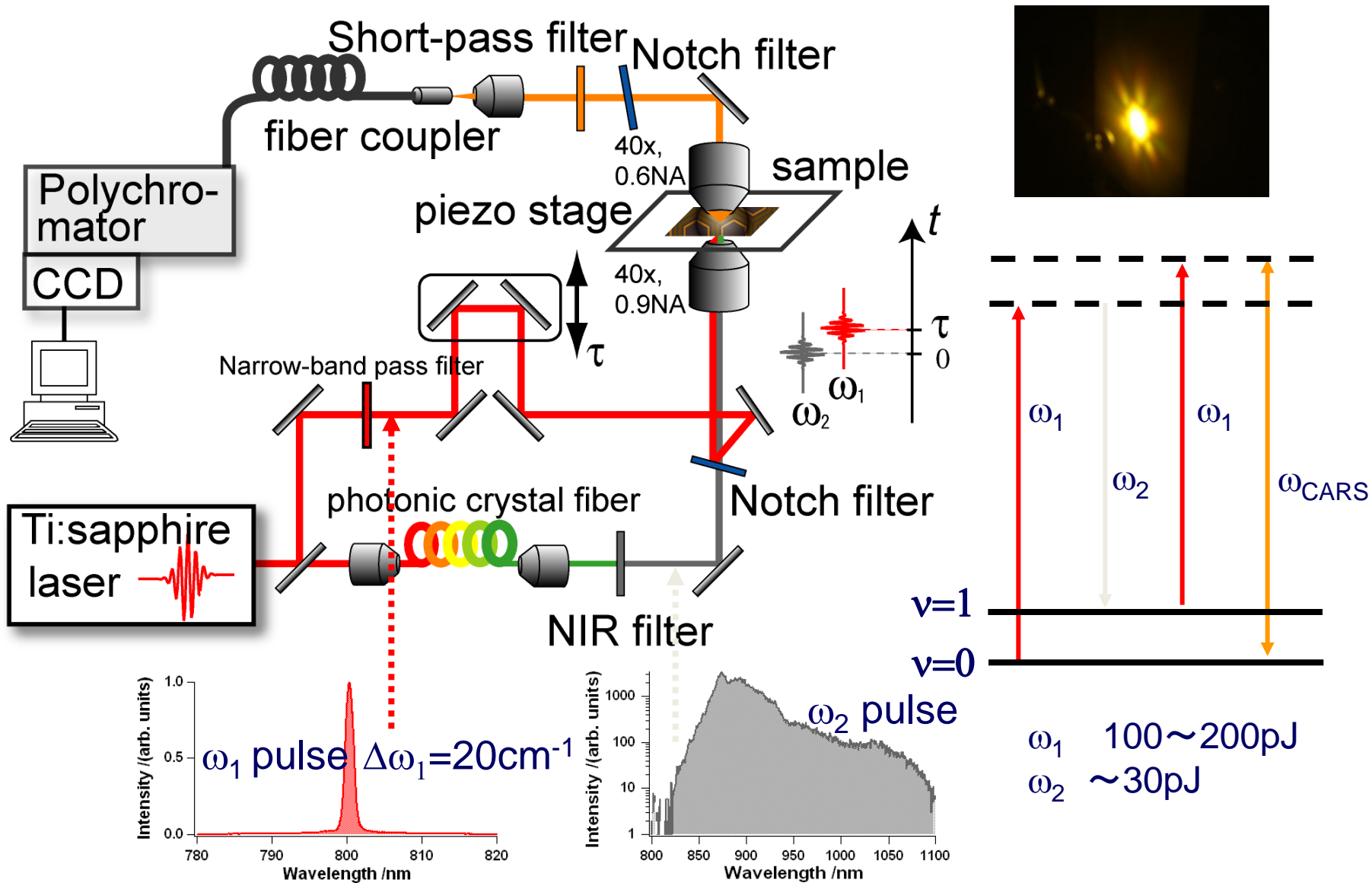


# Organelle Specific Waters in a Living Cell



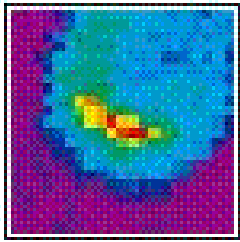
S. Tiwari, M. Ando and H. Hamaguchi, in preparation

# Broadband Multiplex CARS Microspectroscopy

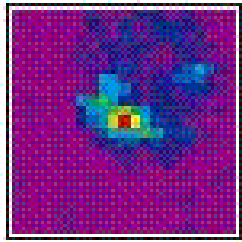


H. Kano and H. Hamaguchi, *Anal. Chem.*, **79**, 8967-8973 (2007).

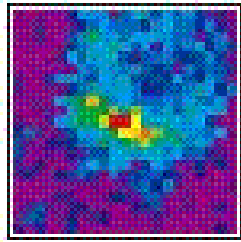
# Vibrational CARS Movies of a Single Budding Yeast Cell



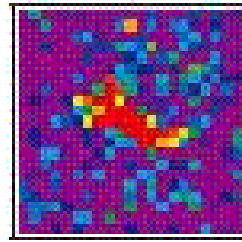
2930 cm<sup>-1</sup>



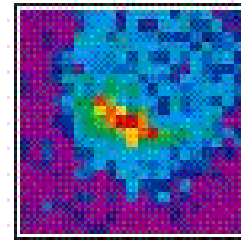
2850 cm<sup>-1</sup>



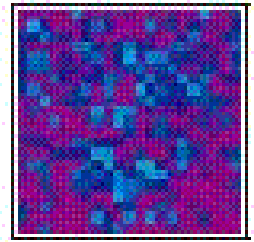
1655 cm<sup>-1</sup>



1602 cm<sup>-1</sup>



1446 cm<sup>-1</sup>



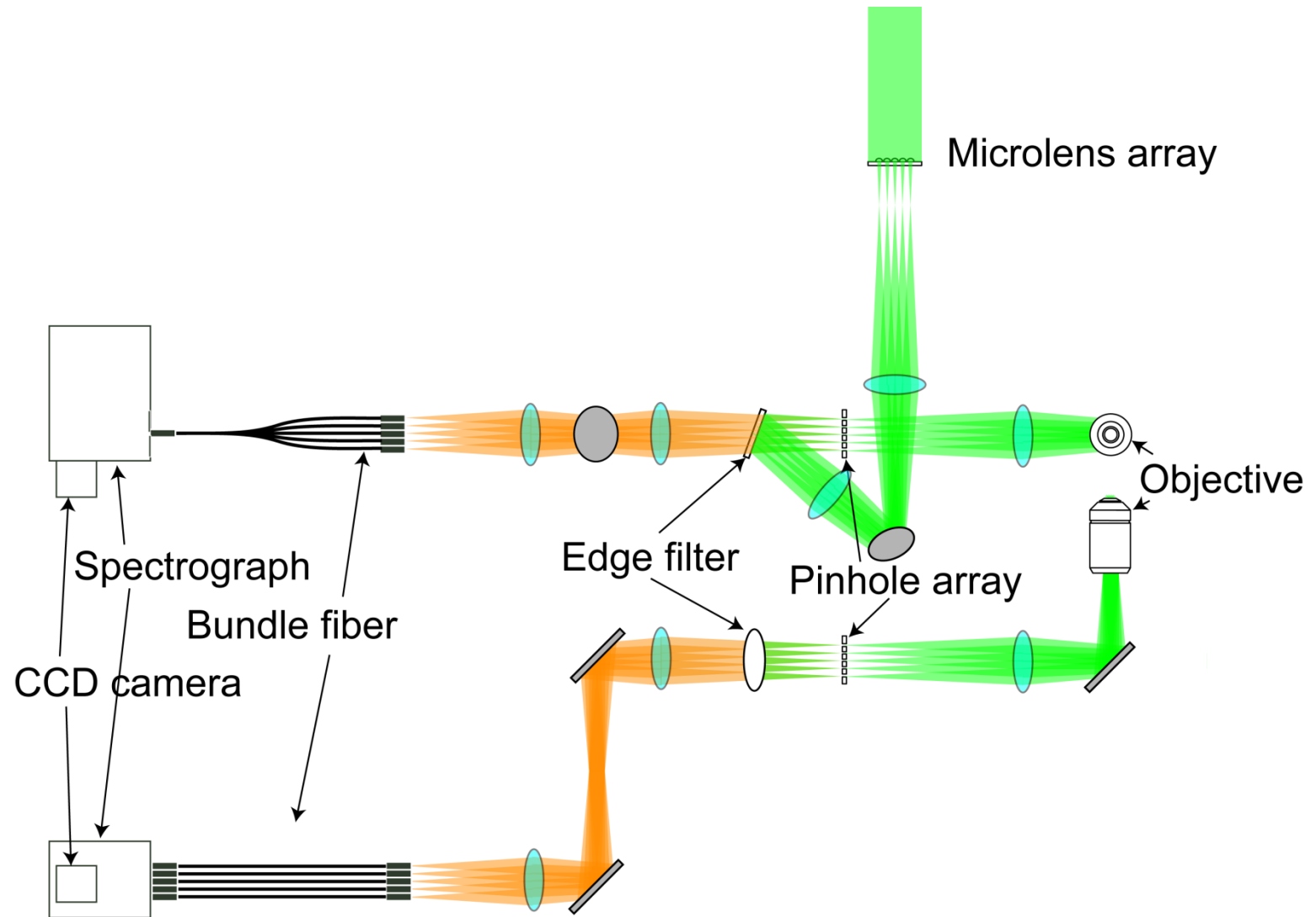
1160 cm<sup>-1</sup>

$\omega_1$ : 10 mW,  $\omega_2$ : 10 mW

Expo. time/pixel: 30 msec

Image acquisition time: ~ 12 sec

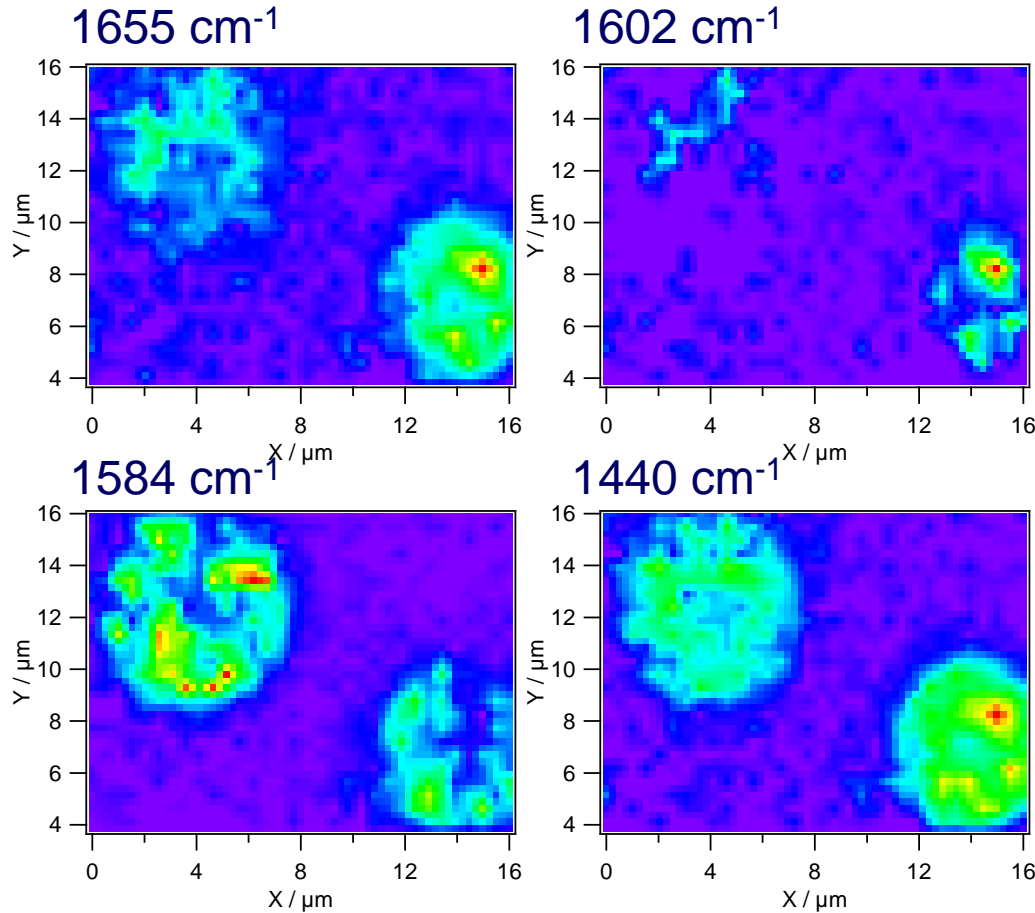
# Multi-focus Confocal Raman Microspectroscopy



M. Okuno and H. Hamaguchi, *Opt. Lett.*, **35**, 4096-4098 (2010).

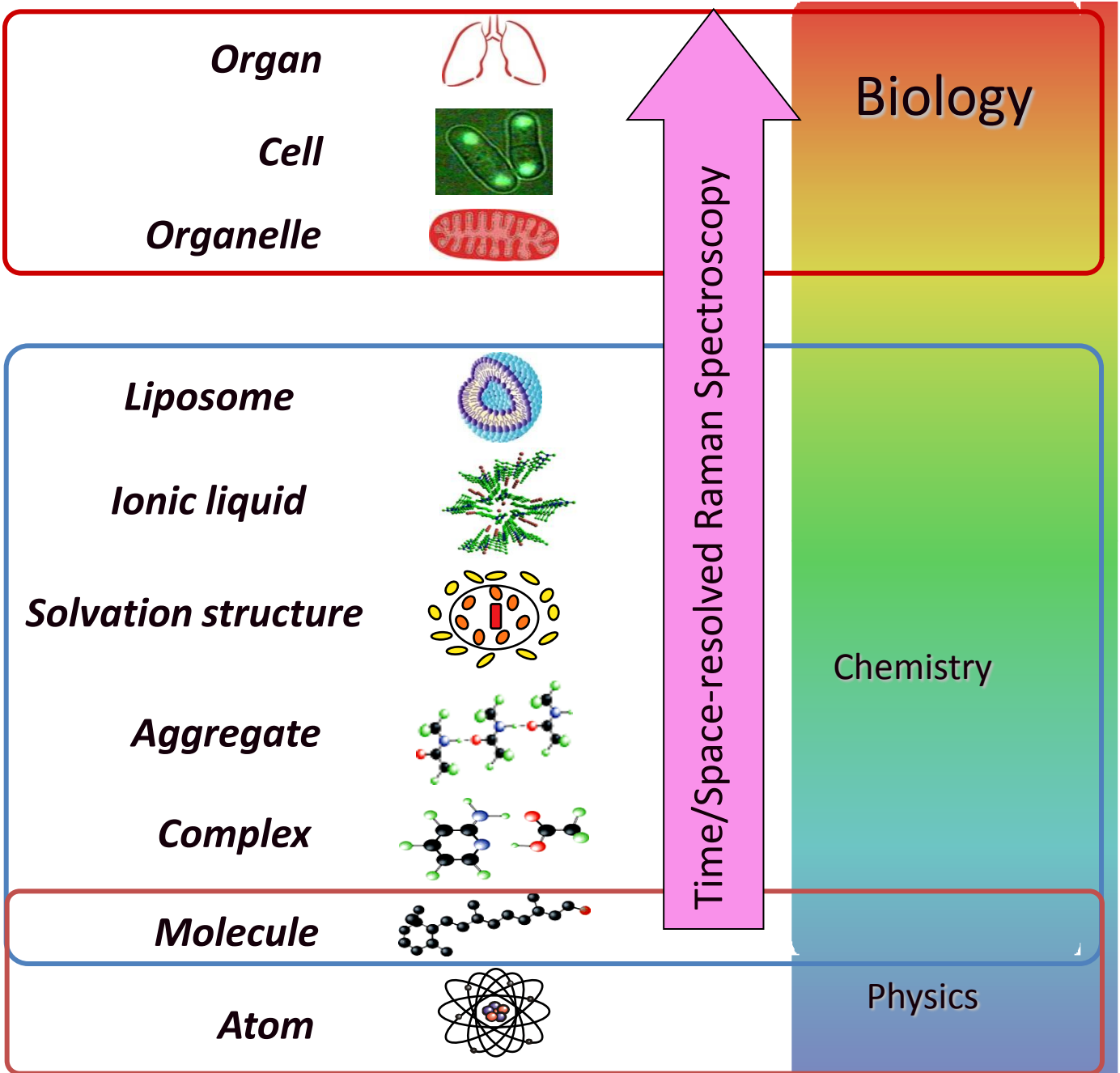
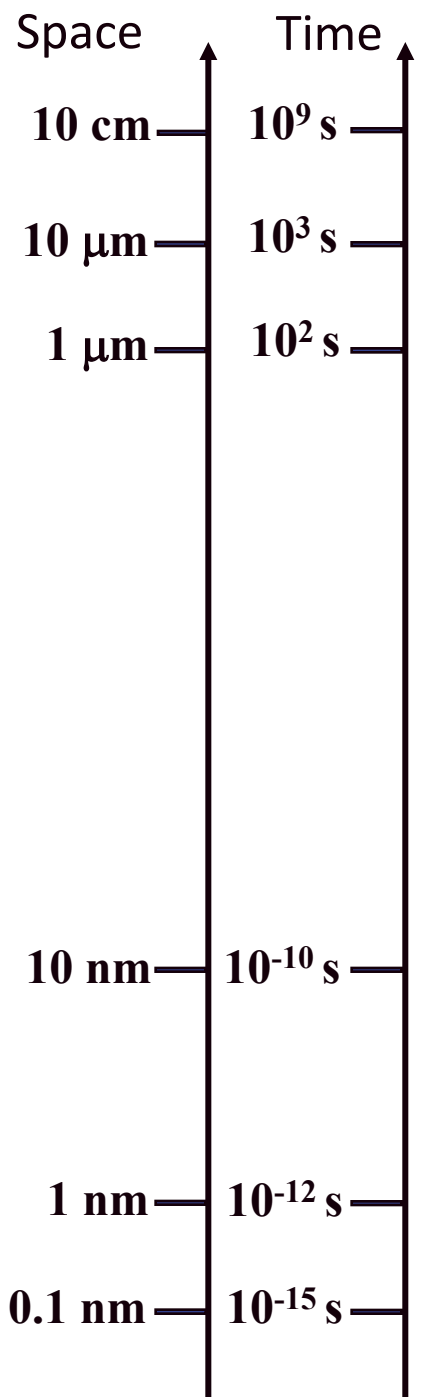


# Budding yeast cells



- ◆ Laser power : 1 mW
- ◆ Exposure time: **1** sec
- ◆ Readout time: ~150 msec
- ◆ PZT scan 4 x 4 points  
2 x 2 μm
- Total area: 16 x 12 μm
- ◆ Total image acquisition time  
(1 sec+ 0.2 sec) x 4 x 4

*~20 sec / image !!*



# Raman Measurement of Food *in situ*



# Raman Application to Food Science



Natural or cultured food resources

**Food processing**

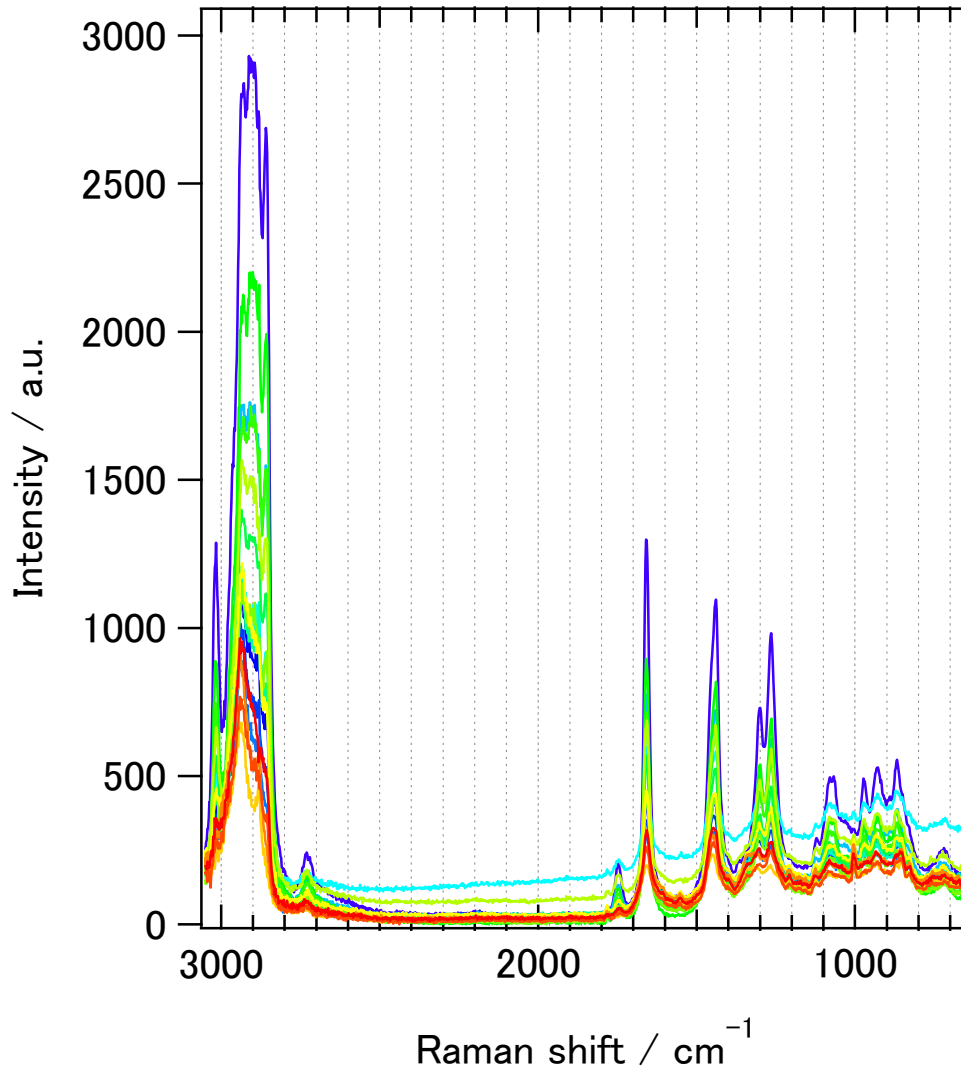


**Food safety & quality control:**

- Food constituent
- Functional ingredient
- Contamination of pathogens or residual pesticides

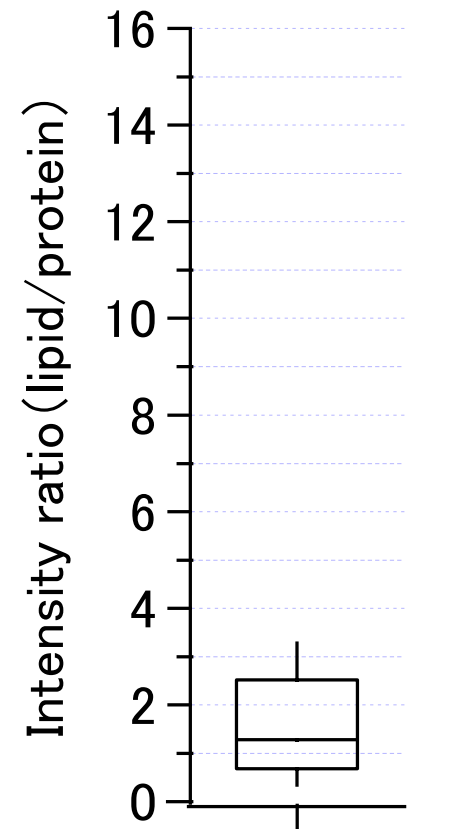
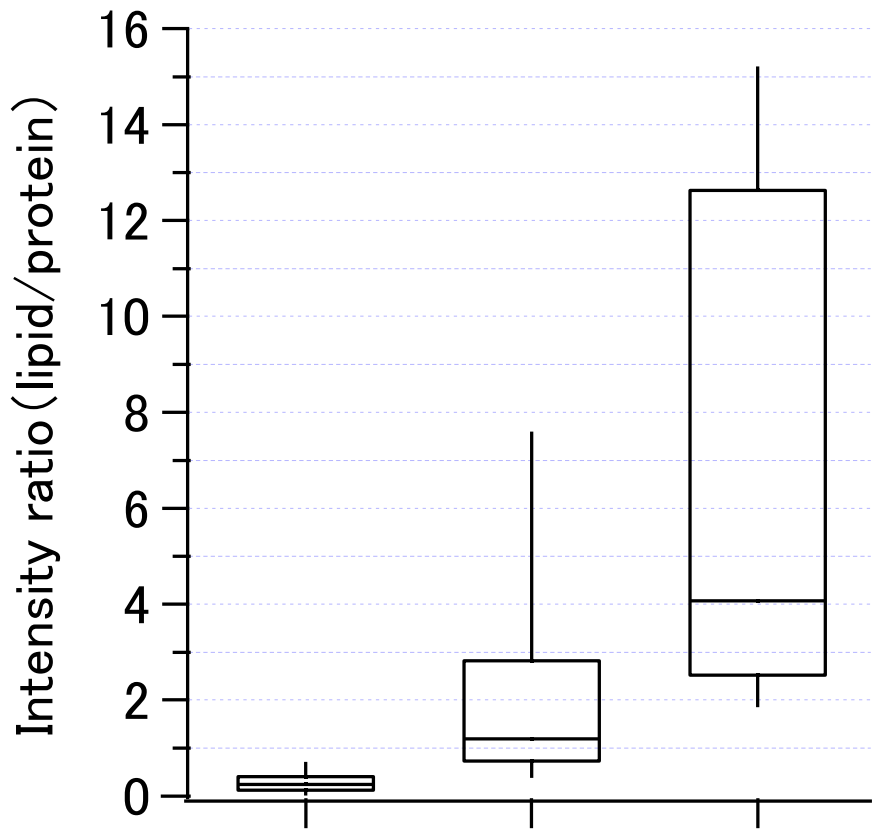
**Label-free, less invasive and rapid analysis  
by using Raman spectroscopy**

# Raman Spectra of Tuna



Quantification of lipids /  
proteins  
⇒ evaluation of food quality  
and taste

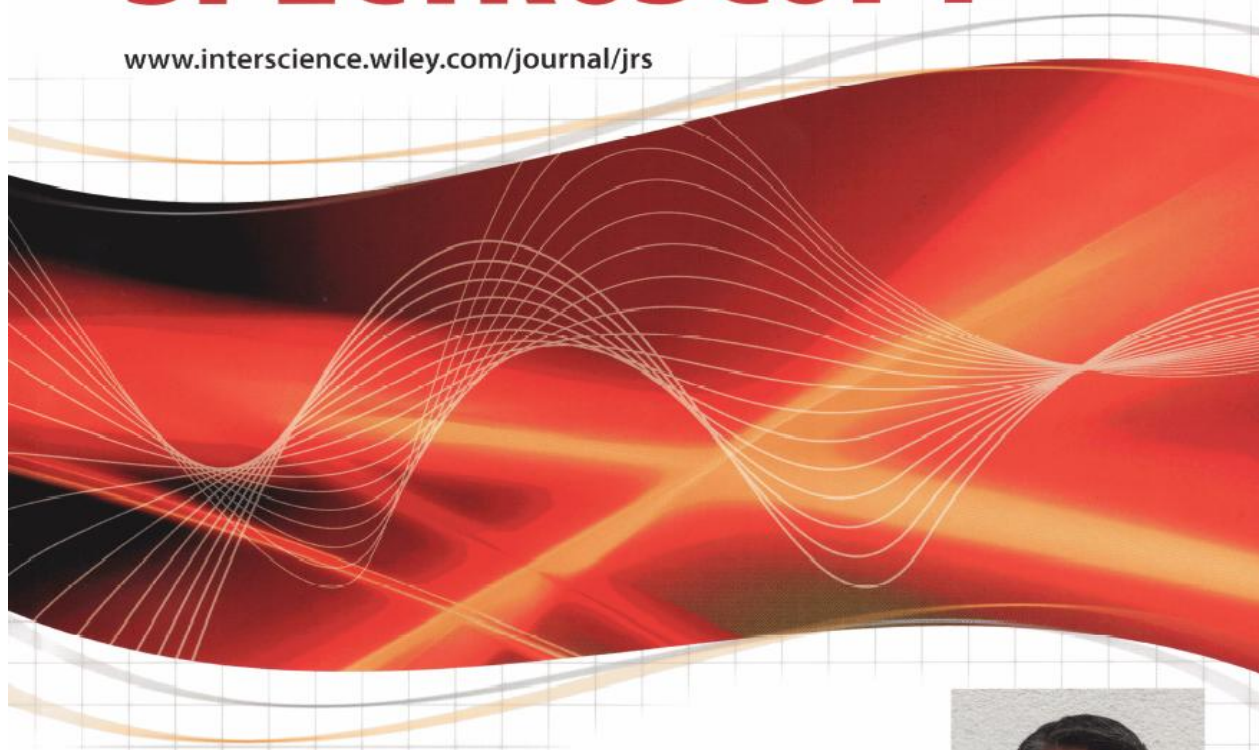
# Lipid Content Analysis of Tuna



Journal of  
**RAMAN  
SPECTROSCOPY**

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[www.interscience.wiley.com/journal/jrs](http://www.interscience.wiley.com/journal/jrs)



**Commemorative Issue:  
For Hiro-o Hamaguchi on the Occasion  
of his 60th Birthday**

Guest Editors: V. Deckert, M. W. George and S. Umapathy



 **WILEY**

# First Student Summer Camp of Taiwan Association of Raman Spectroscopy (2013.7.6 at Jianshi)

